

Microscope Basics

Midlands Cryo-EM Workshop
2021

Christos Savva

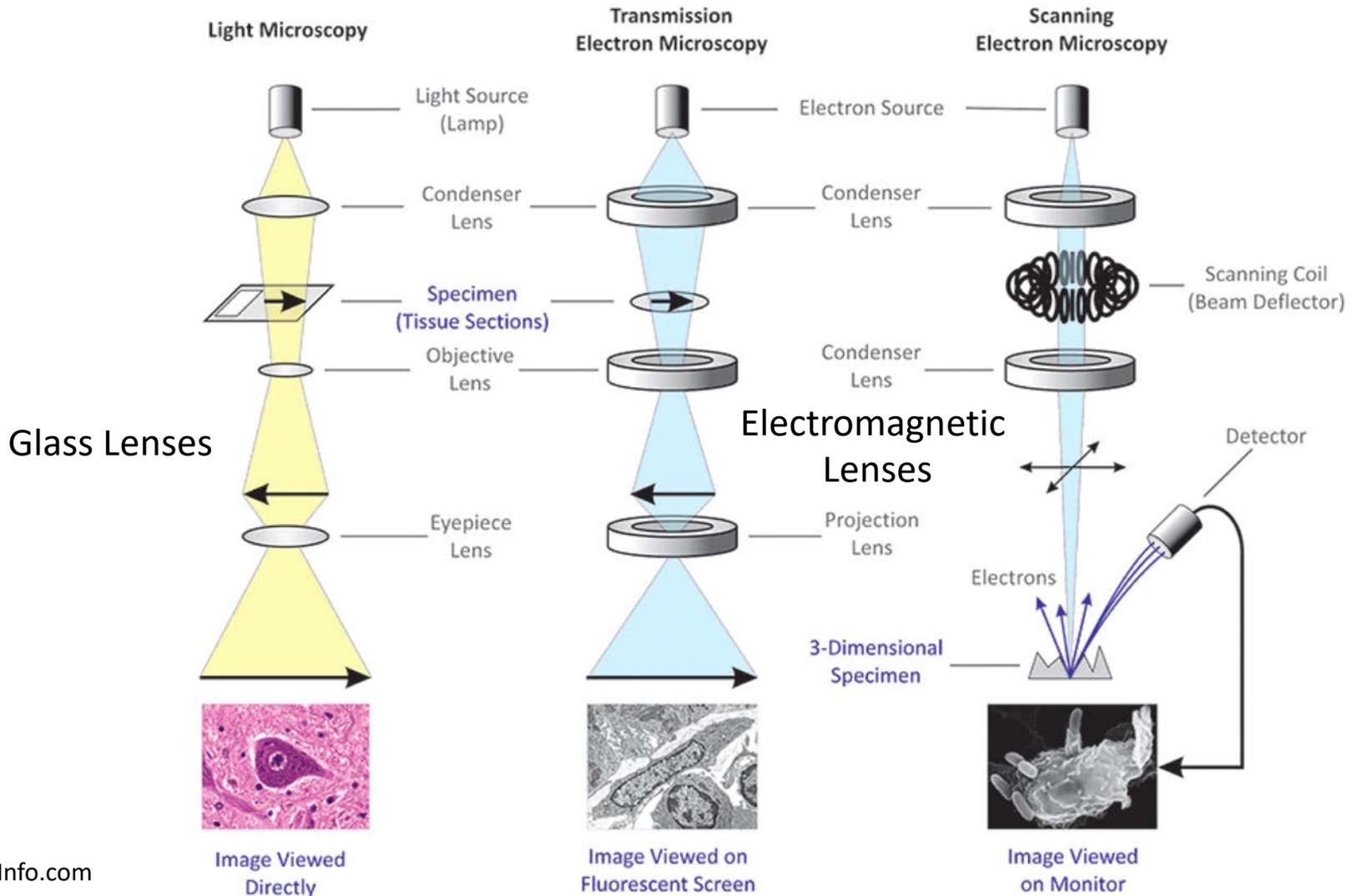


1931



2021

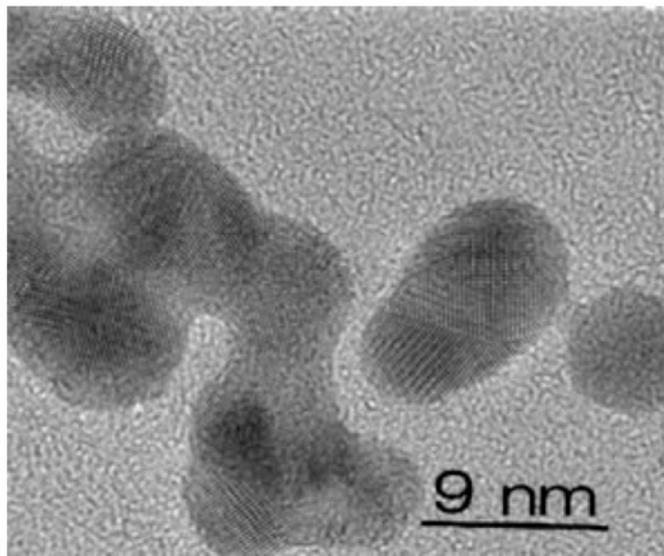
Electron Microscopy



Transmission Electron Microscopy

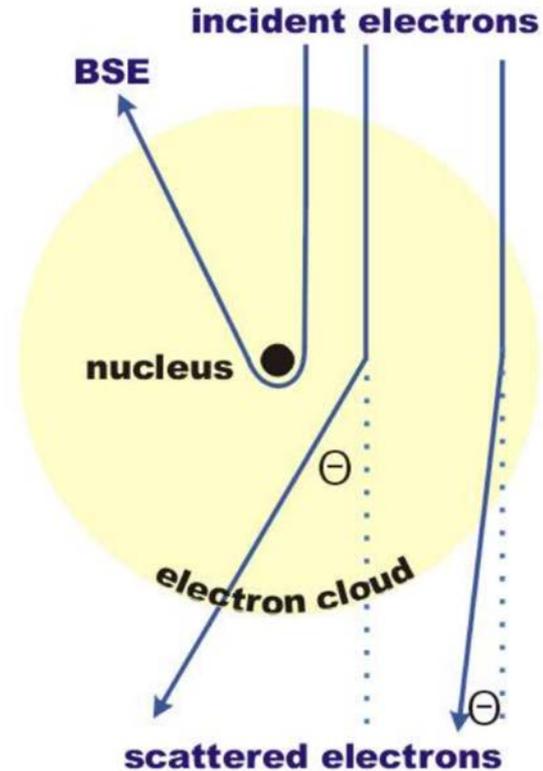
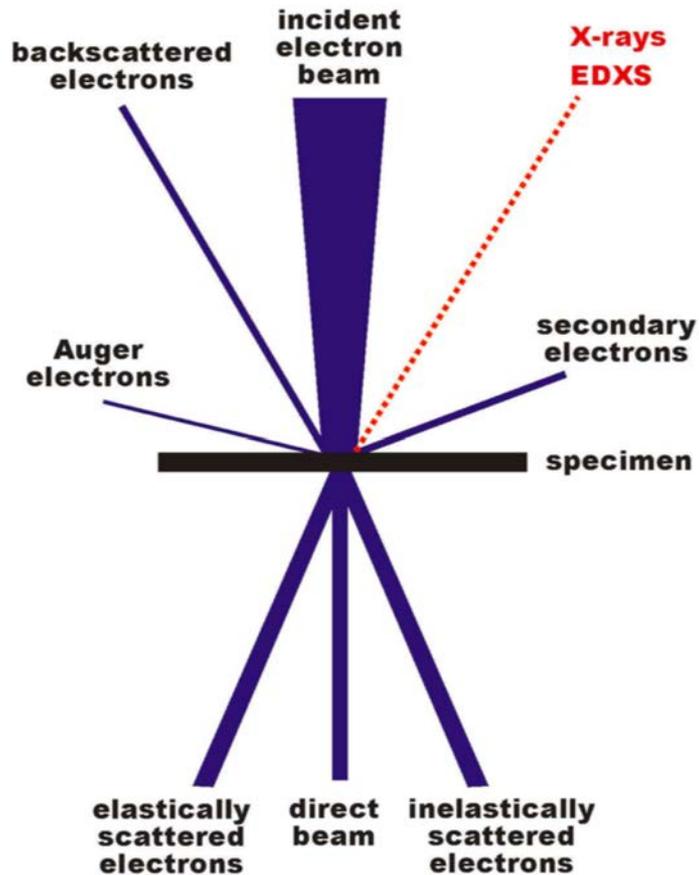
- The Transmission electron microscope is required for high-resolution structural studies
- Requires ultra-thin specimens $<0.2 \mu\text{m}$
- 100 kV electrons $\lambda=0.037 \text{ \AA}$
- Lens aberrations limit resolution to $\sim 1 \text{ \AA}$ (0.5 \AA in some cases)

Electron Micrograph: Picture taken with an EM



Gold particles on carbon film. Lattice spacing of 2.04 \AA

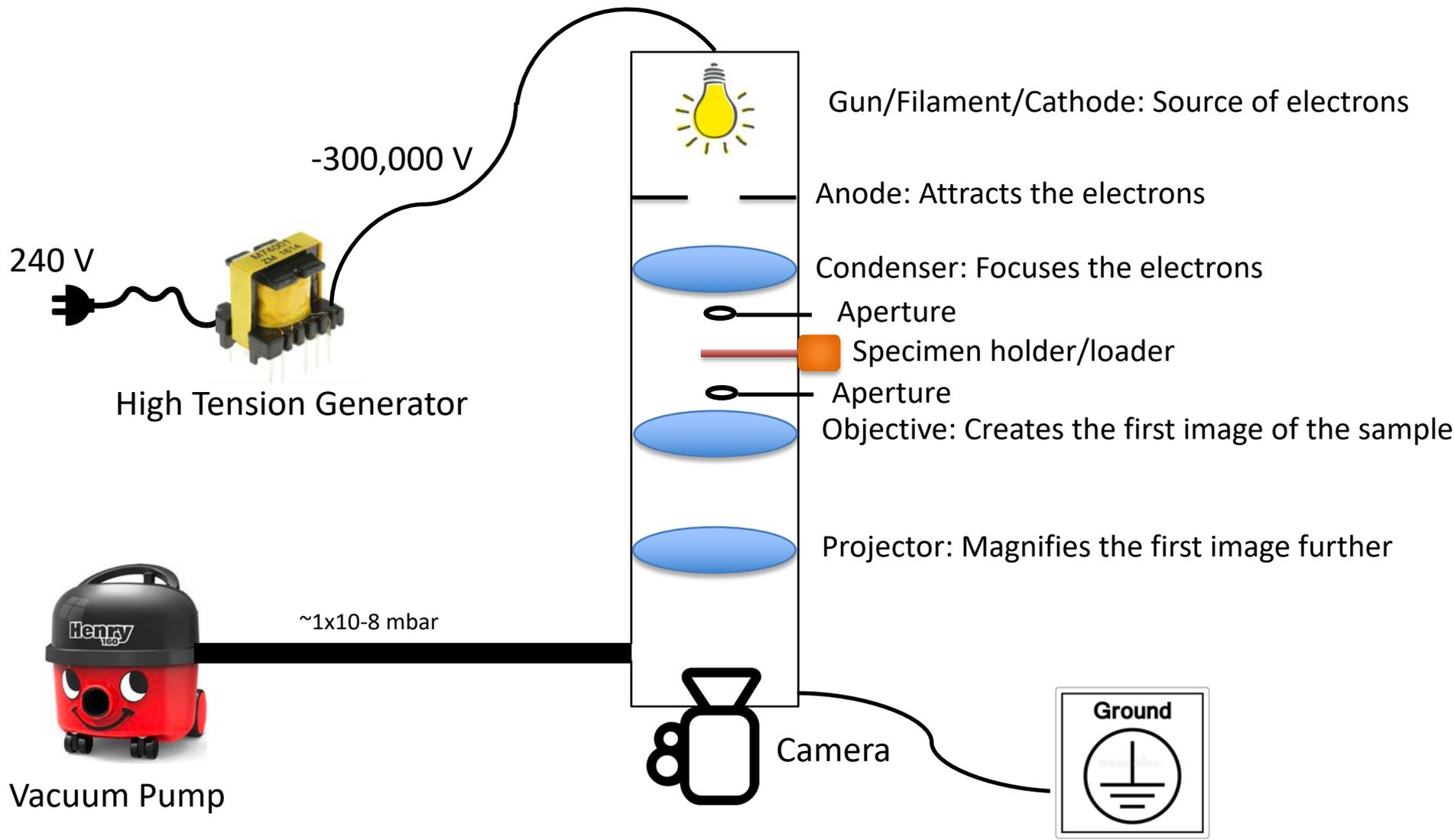
Transmission Electron Microscopy



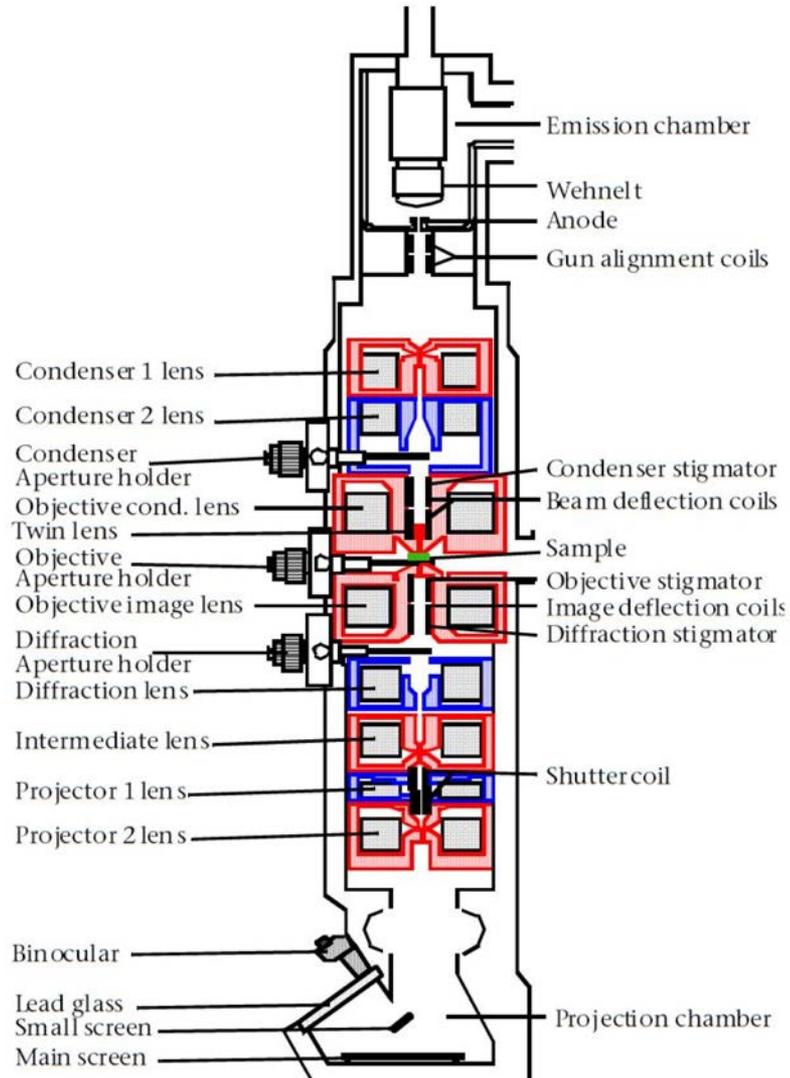
Electrons attracted to nucleus of atom. The higher the atomic number (Z) the higher the scattering.

A very Basic TEM

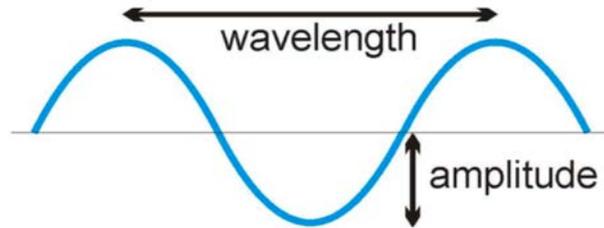
(that probably wont work)



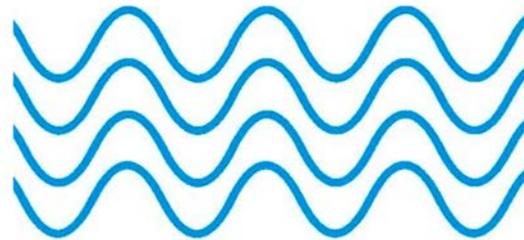
The Transmission Electron Microscope



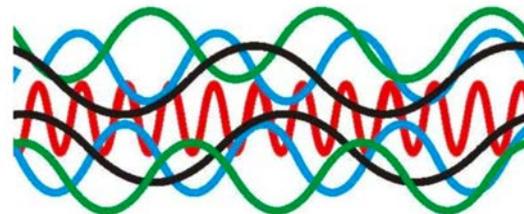
Wave Coherence



- Spatial Coherence: Are the electrons coming from the same direction and in phase?
- Temporal coherence: Do the electrons have the same energy/speed (wavelength)?



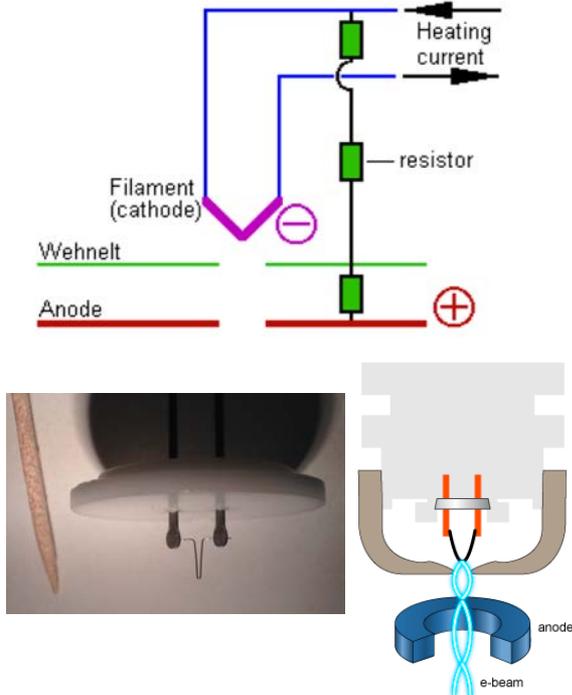
Coherent Waves



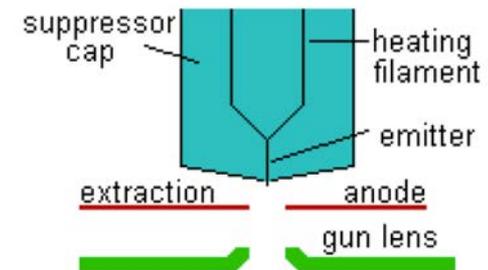
Incoherent Waves

TEM Gun types

Thermionic: W or LaB6



Field Emission Gun (Schottky)

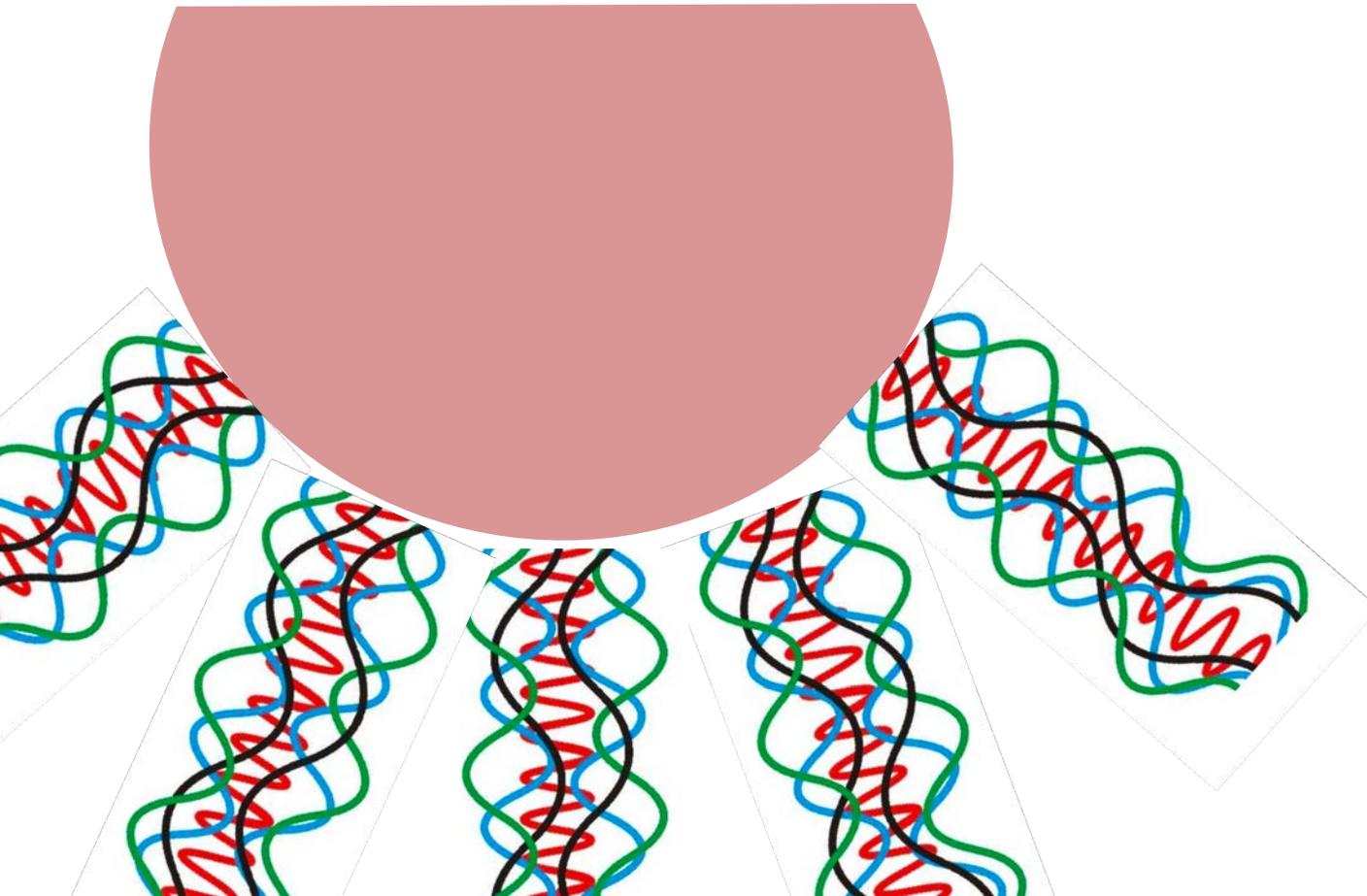


100nm

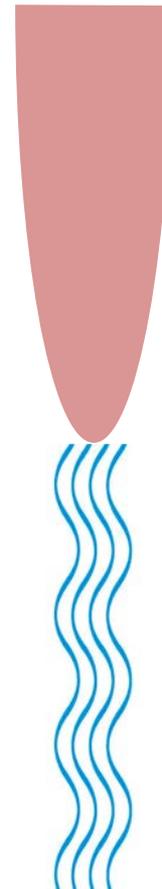
Temporal	1.0-2.0	Energy Spread (eV)	0.2-0.3
Spatial	$>10^4$	Source Size (nm)	<30
	10^5	Brightness ($A/cm^2 \cdot xsr$)	10^8
	10^{-3}	Vacuum required (Pa)	10^{-8}
	100-300	Life Span (hrs)	~ 8000

Electron Beam Coherence

Thermionic gun

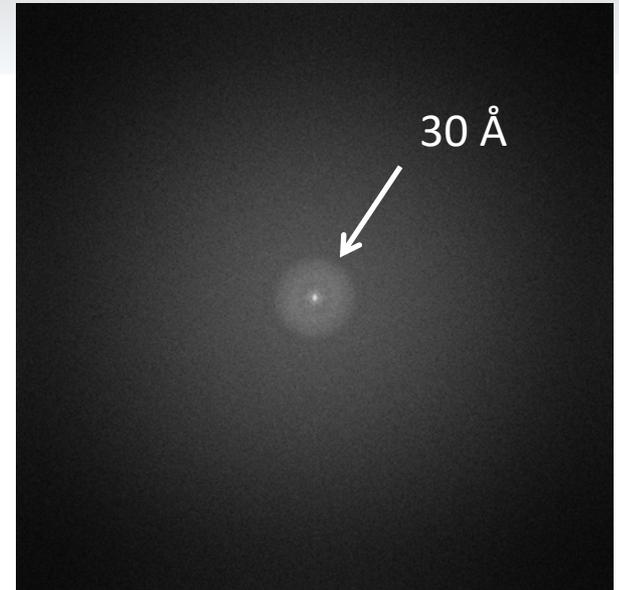
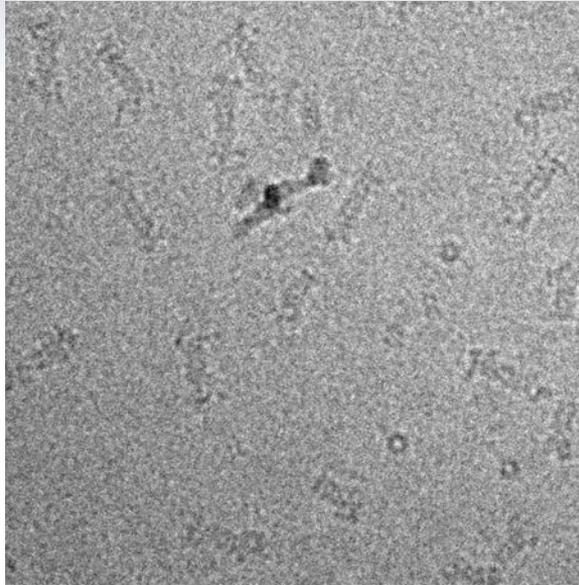


Field Emission Gun

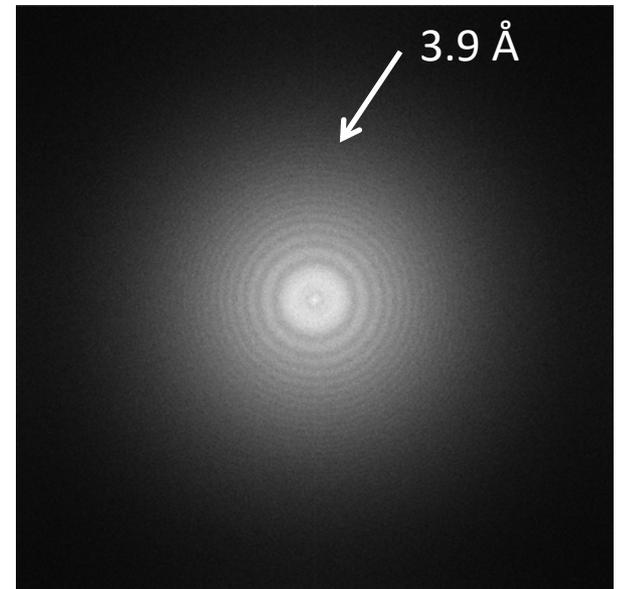
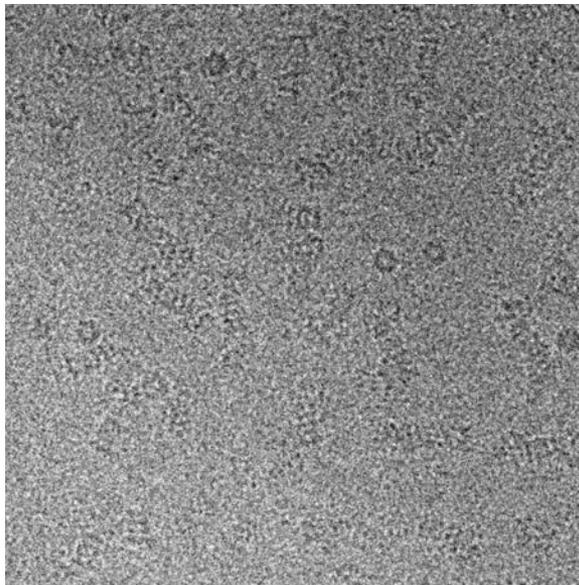


Gun Dependent Resolution

Thermionic Gun

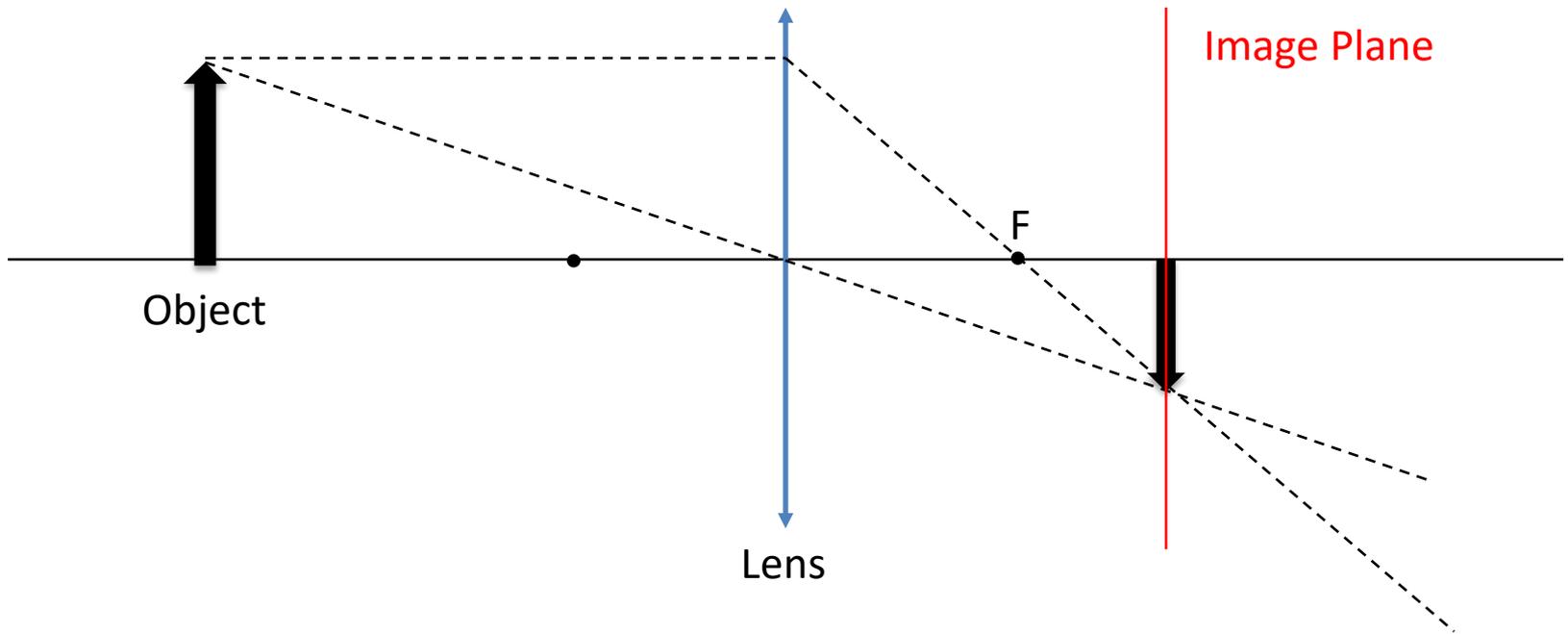


Field Emission Gun



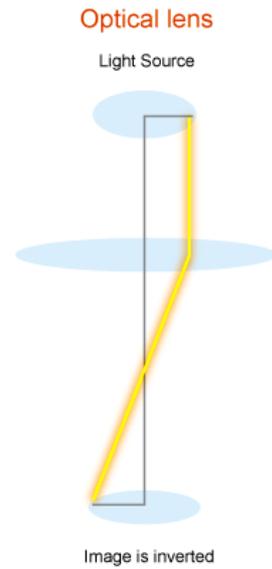
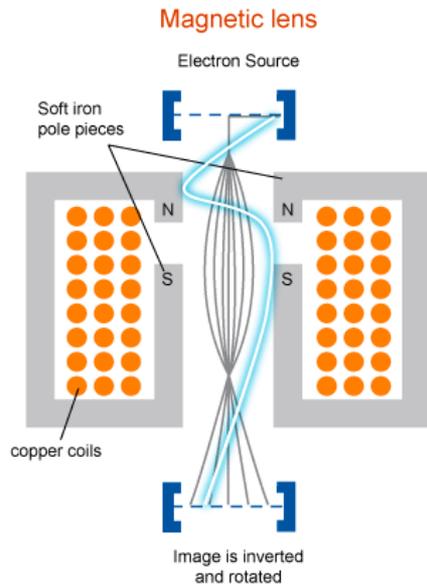
(Alice Clark, 2014)

Lenses



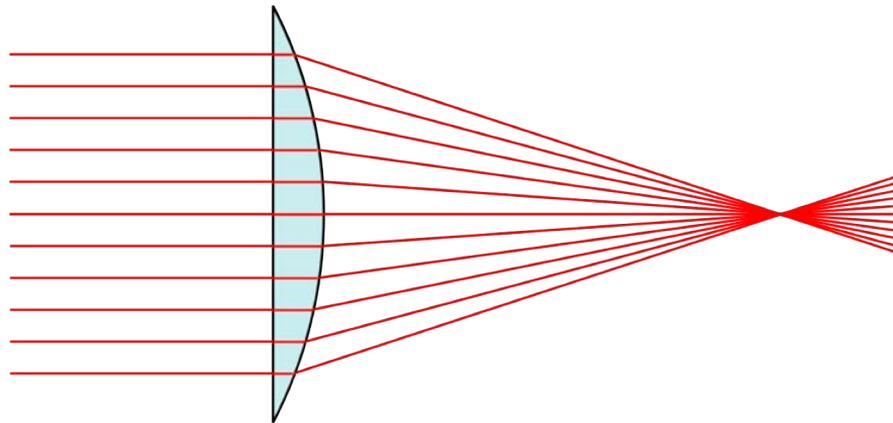
F=Focal point

Lenses

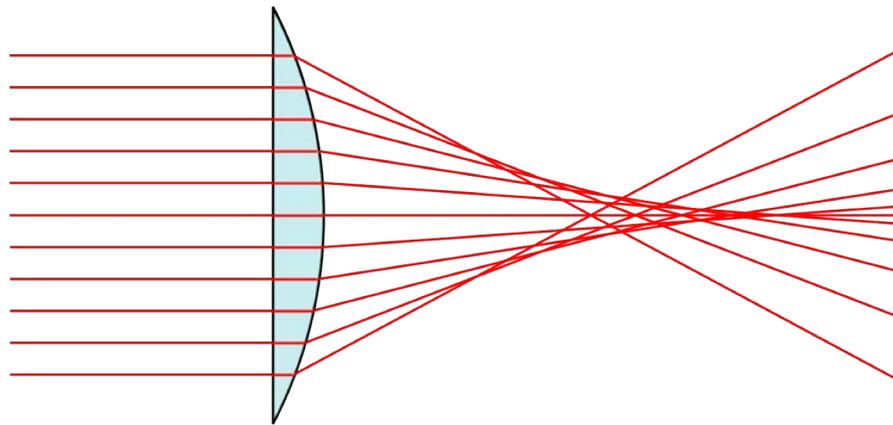


Lens aberrations

Lenses are not perfect and suffer from aberrations



Perfect Lens

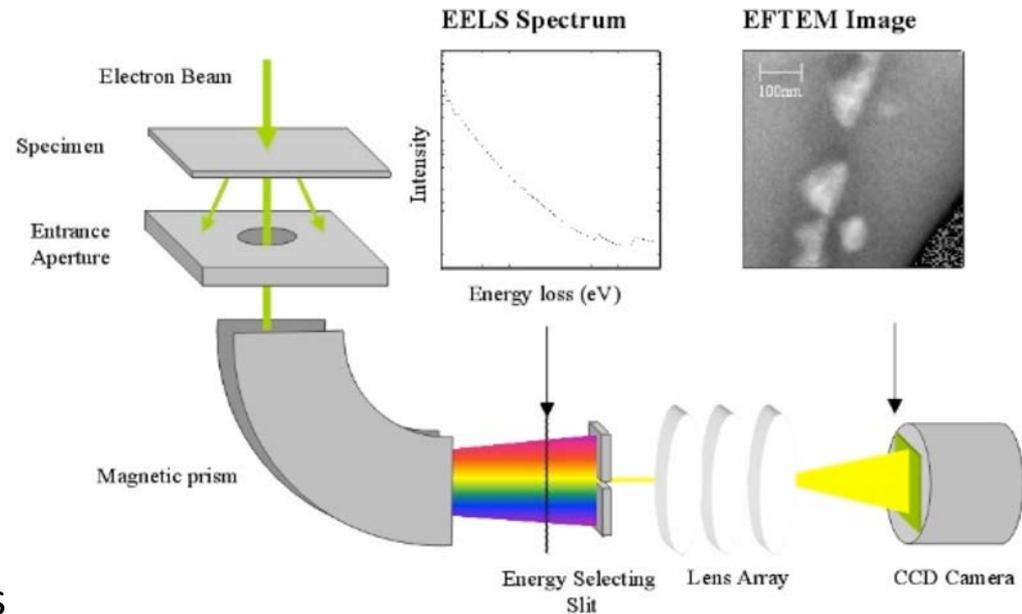
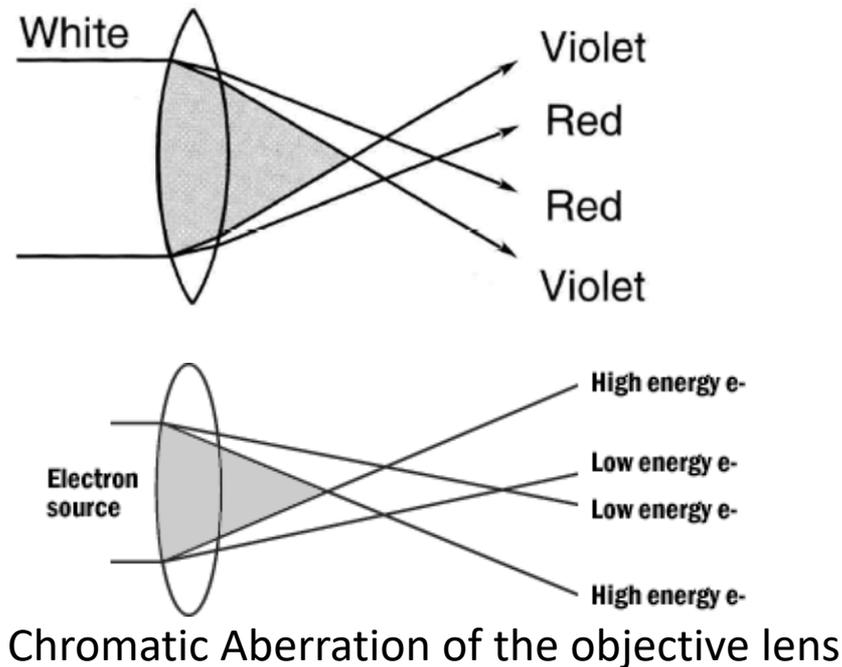


Spherical Aberration (C_s)

Requires expensive hardware to correct

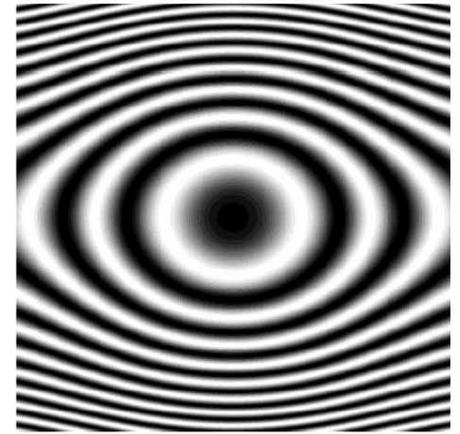
Chromatic aberration (C_C)

- Energy spread of electrons due to gun temporal coherence
- Thick samples also result in many electrons with lost energy
- Cs correctors available but expensive
- Energy filters can remove energy loss electrons
- More important for tomography of thick specimens

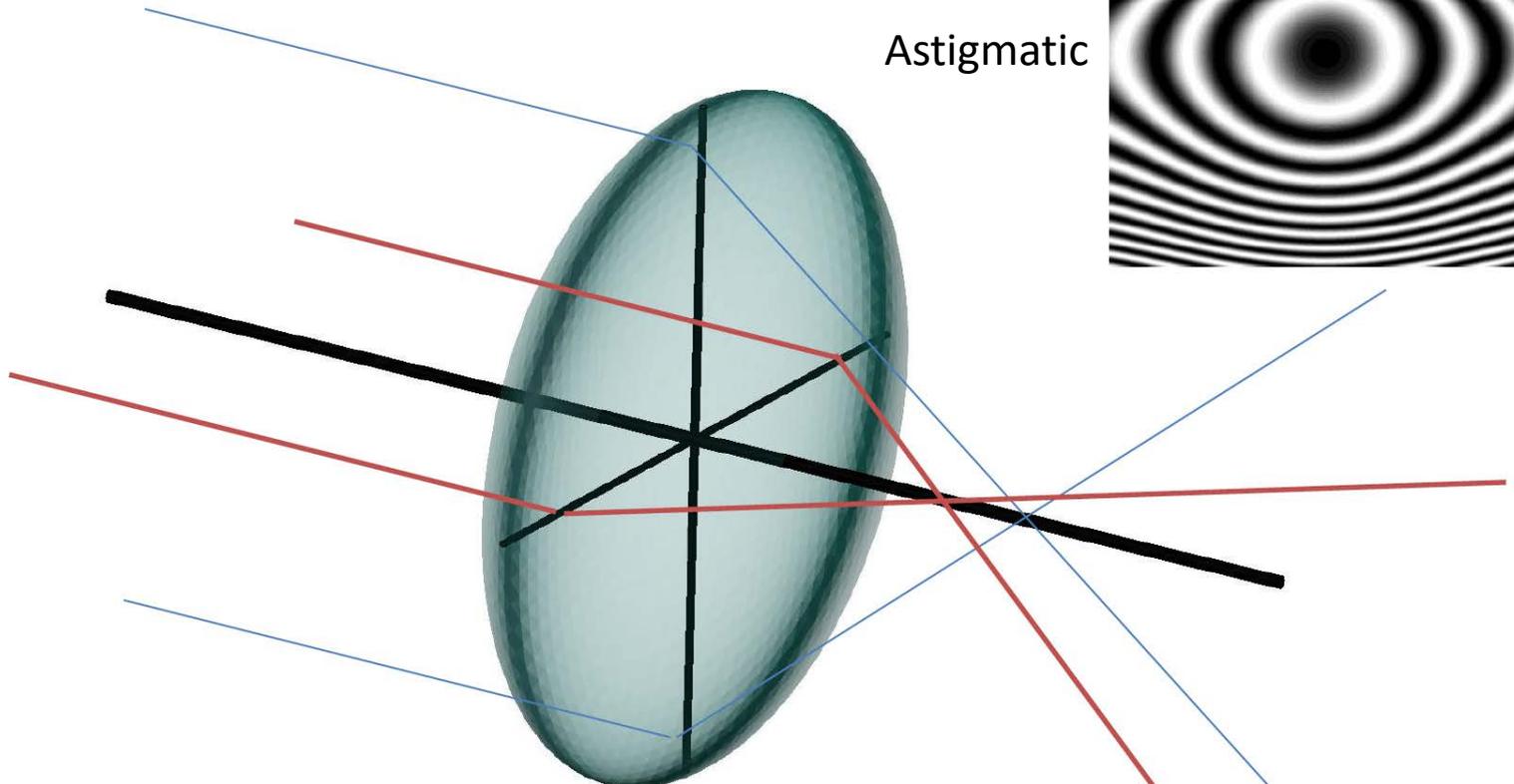


FFT of sample

Objective Lens Astigmatism

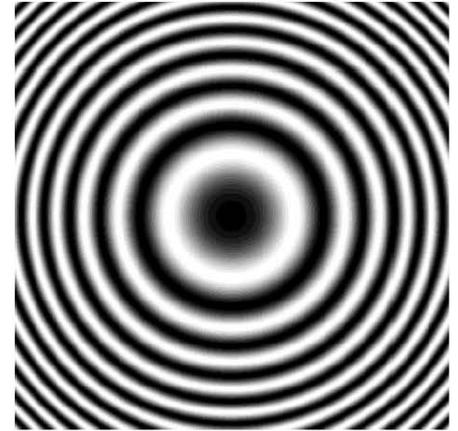


Astigmatic

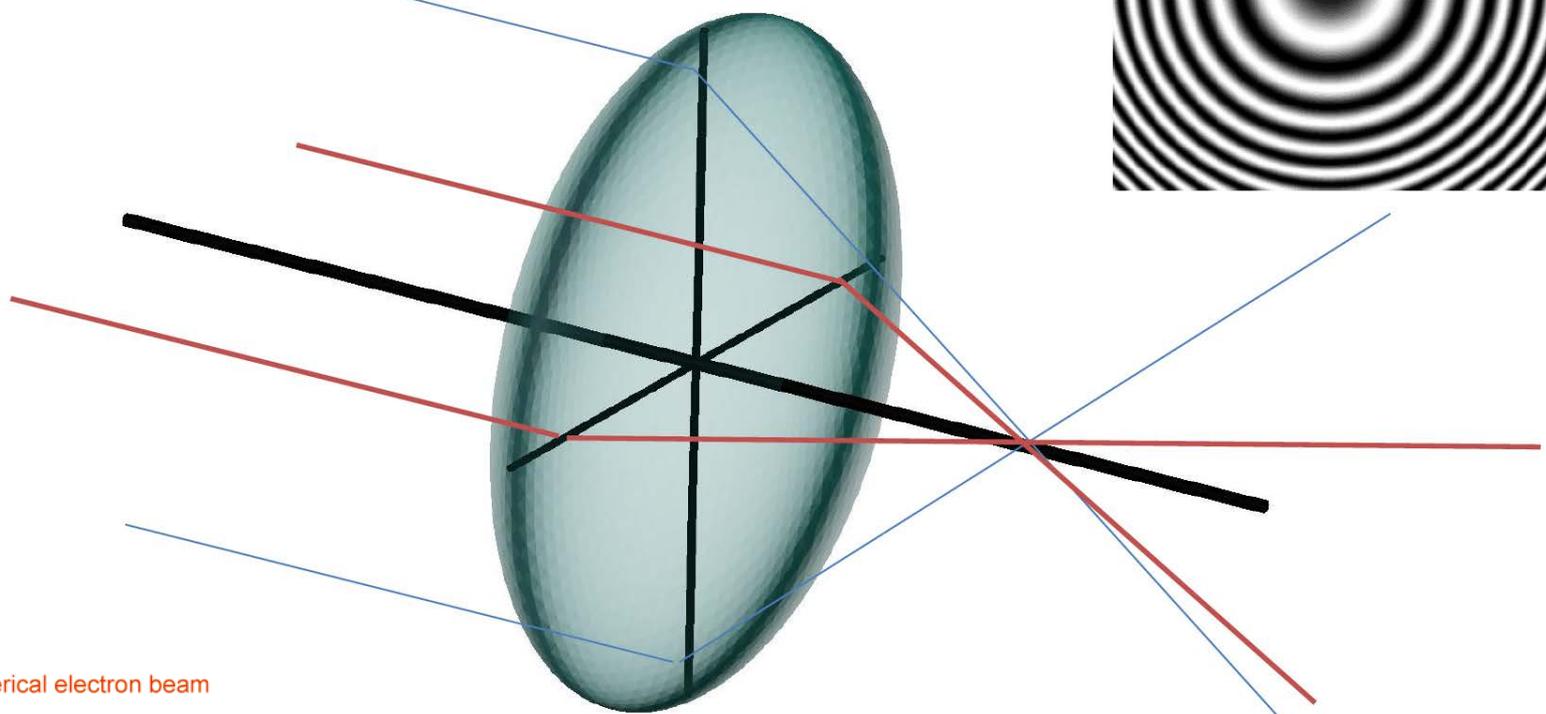


Astigmatism

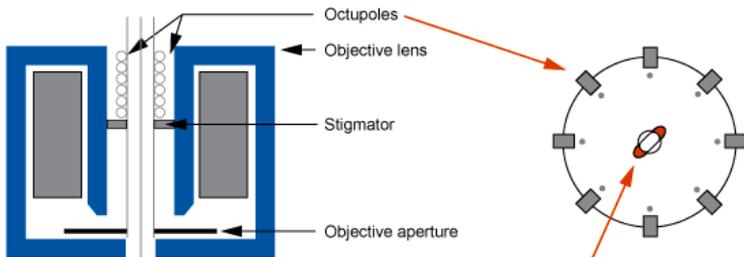
Astigmatism is easy to correct on modern microscopes



Corrected



Astigmatism: Non-spherical electron beam

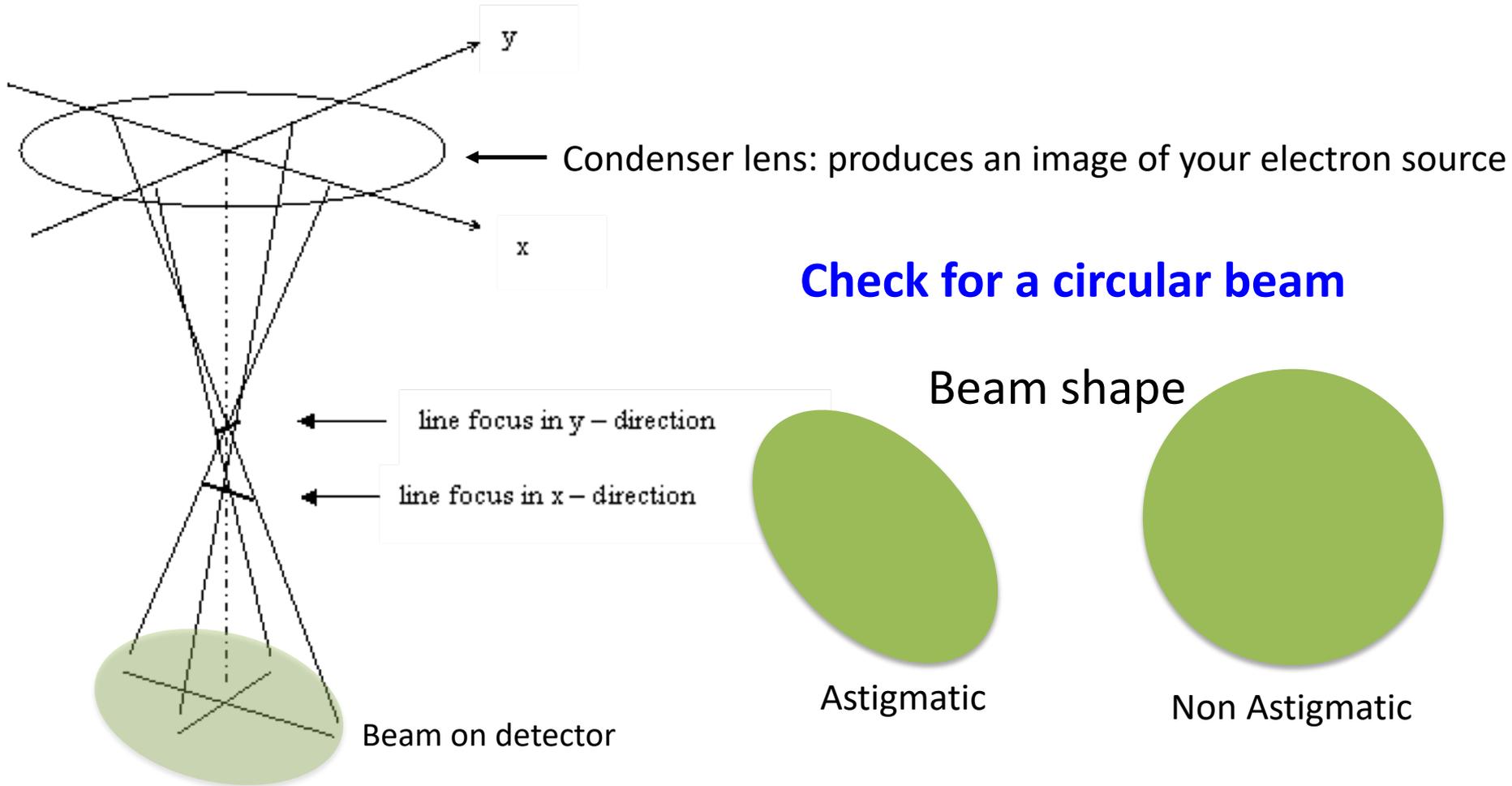


Astigmatism: Beam

Astigmatism

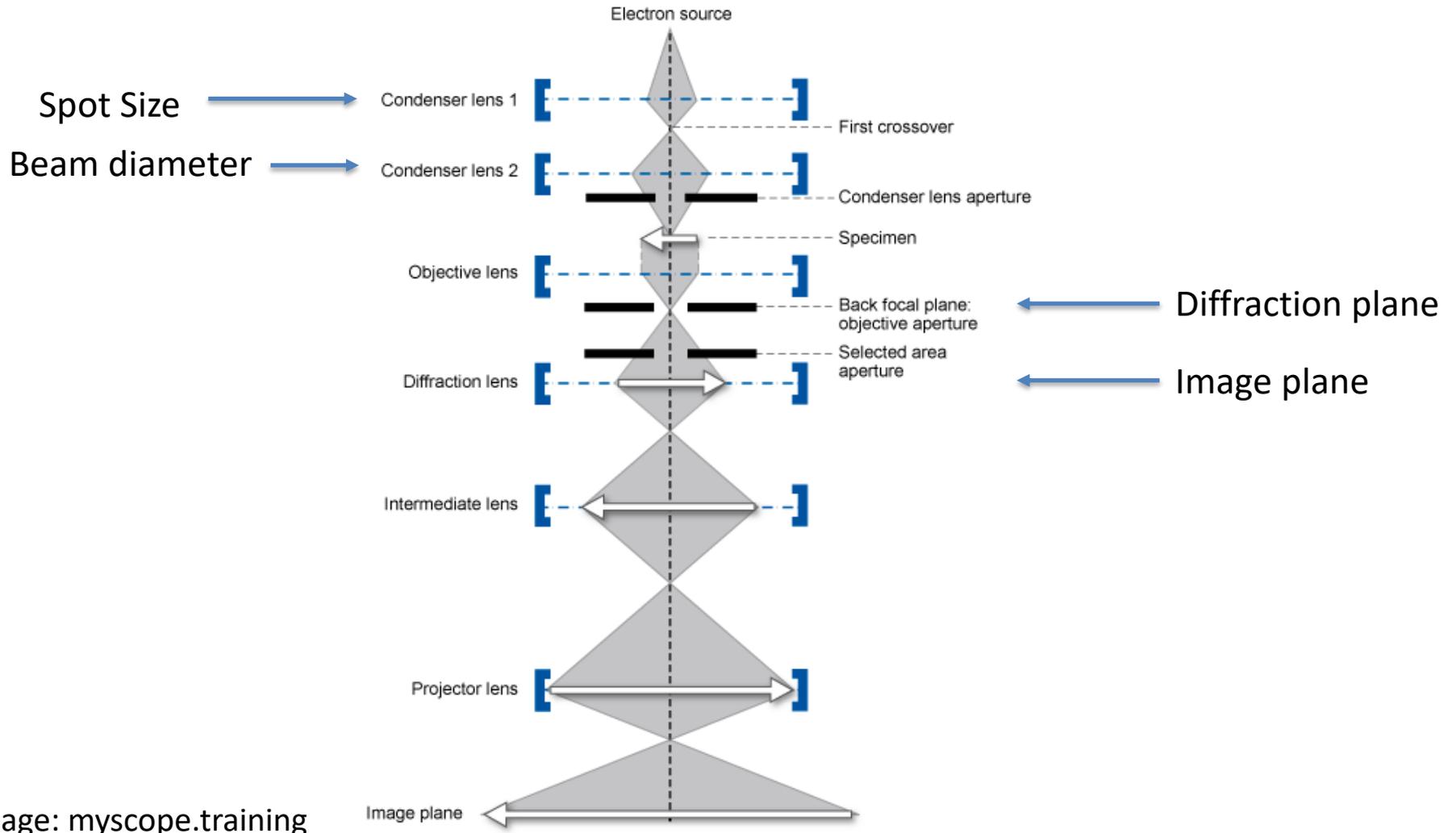


Condenser Lens Astigmatism



Electron beam path through the lenses

Microscopes with 2 condenser lenses



Choice of Accelerating Voltage

What kV should I use? What to consider:

- Resolution: Wavelength at 300 kV vs smaller than at 100 kV
- Aberrations: higher effect of lens aberrations higher at lower kV
- Radiation damage: 1.5 x less damage at 300 kV vs 100 kV
- Useful information: 25% higher elastic/inelastic ratio at 100 kV
- Detector performance: DDD perform worst at lower kV.
Detectors for lower kV scopes have reduced field of view
- Cost: 100 kV much cheaper than 200 or 300 kV
- Sample thickness: Can image thicker samples at higher kV

A 100 kV FEG TEM



Tundra Cryo-TEM

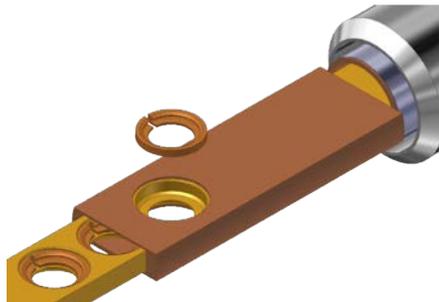
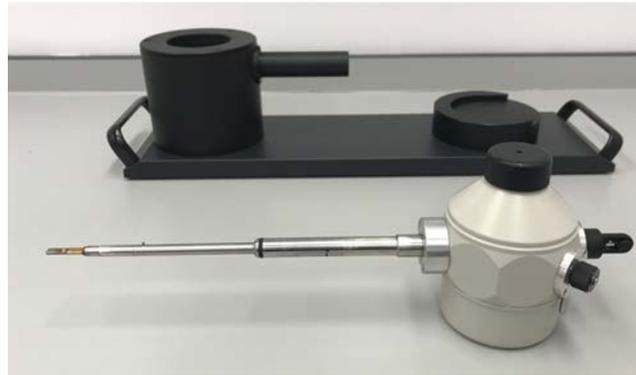
Specimen Holders: Side entry

Samples kept at $\sim -175^{\circ}\text{C}$

Dubochet MKI



Gatan 626: 1 grid



Gatan 910: 3 grids at once



Simple Origin 200: Takes 2 autogrids

Specimen Holders: Side entry

Advantages

- Cheap (£40-80K...yes that's cheap!!)
- Can be used on almost any side-entry TEM (Anti-contaminator required)
- Was the only option until recent years

Disadvantages

- Fiddley to use
- Fragile/easy to break
- Require pumping/heating for optimal performance (Pumping station and Heater)
- Low throughput (Sample exchange takes ~45-60min)
- Each load cycle introduces moisture to the column
- Stability not great: Drift, vibration prone
- Require manual LN2 top-up

Autoloaders

TFS Glacios 200 kV



TFS Krios 300 kV



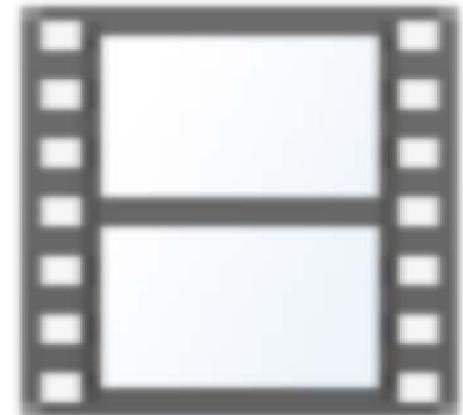
JEOL Cryo-ARM 200



JEOL Cryo-ARM 300



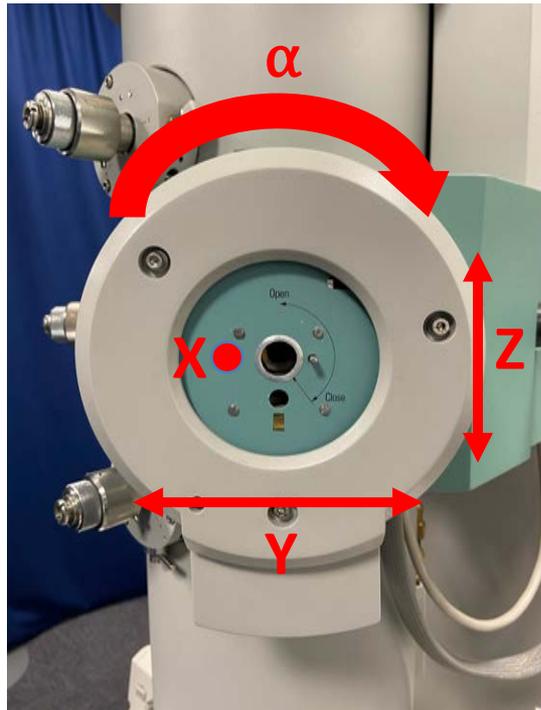
Sample preparation
for TEM



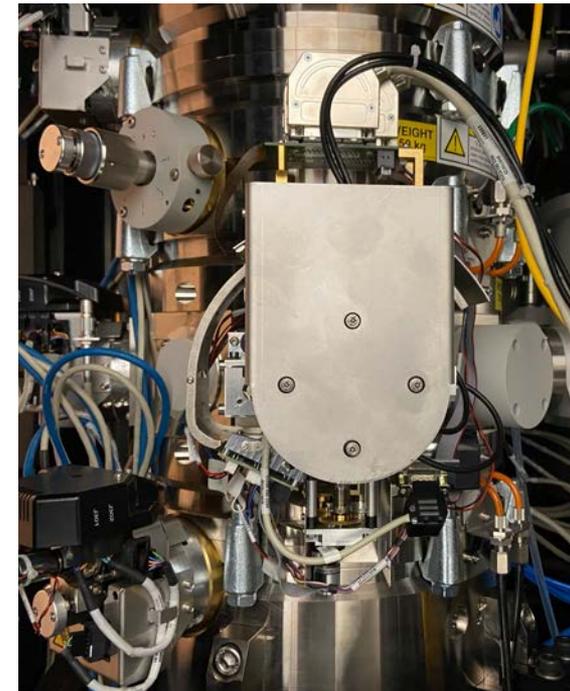
Microscope Stage

- The stage or goniometer supports the specimen holder
- On Autoloader systems the holder is always inserted
- Moves along X, Y and Z directions and tilts along α (and does the same to your sample)

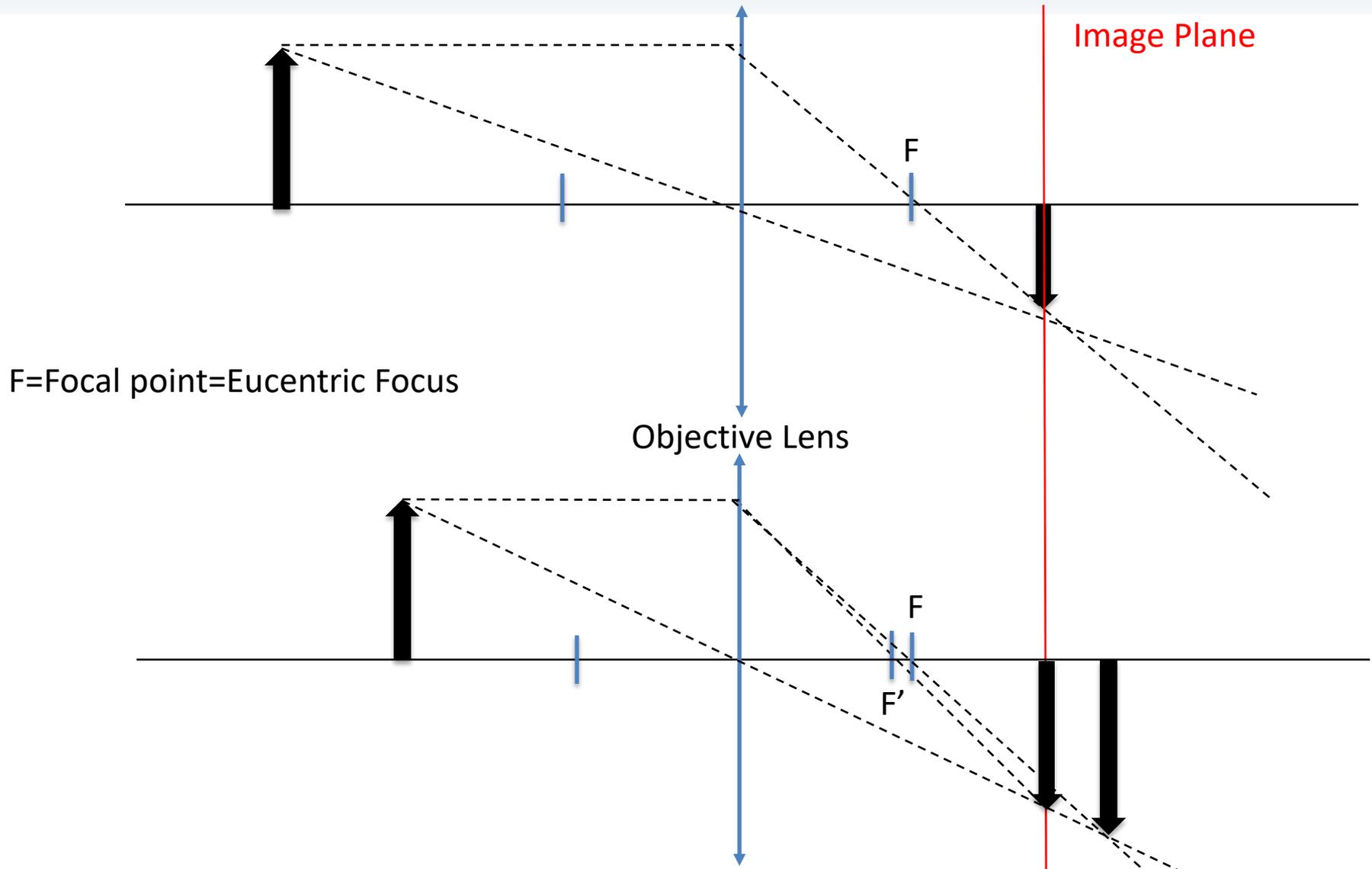
Philips CM200 Stage



Krios Stage



Setting the sample to Eucentric Position



Detectors for the TEM

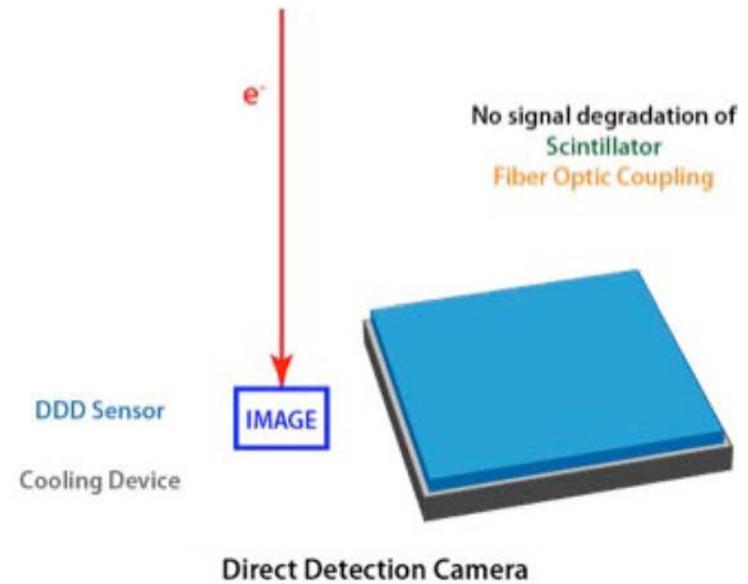
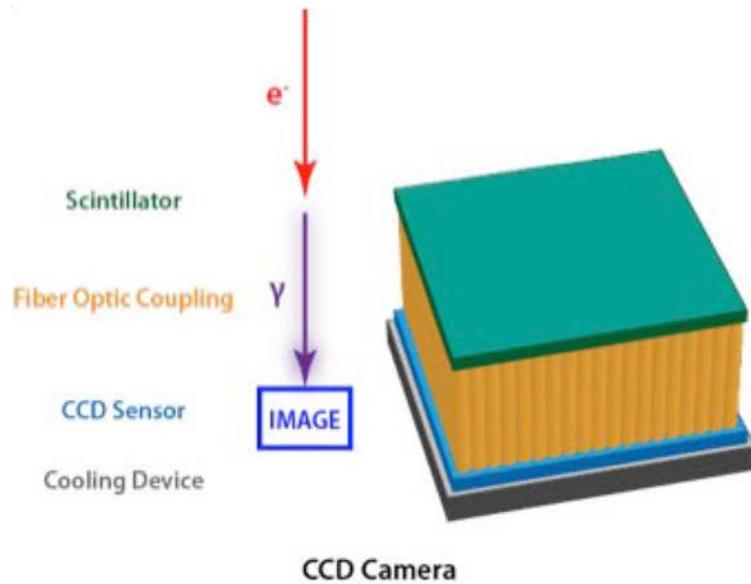
Detective Quantum Efficiency (DQE) = SNR_o^2 / SNR_i^2

A measure of the signal to noise ratio degradation

Perfect detector has DQE of 1

Detector	Advantages	Drawbacks
Film	<ul style="list-style-type: none">- Large area- Decent DQE	<ul style="list-style-type: none">- Limited to 50 exposures- Needs developing-Scanning- Adds moisture to microscope
CCD	<ul style="list-style-type: none">- Easy to use- Instant	<ul style="list-style-type: none">- Low DQE (0.1)
Direct electron Detectors	<ul style="list-style-type: none">- High DQE- Fast frame rates-Movies	<ul style="list-style-type: none">- Expensive

CCD vs DED

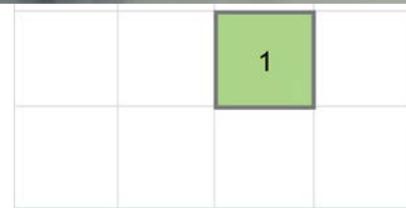
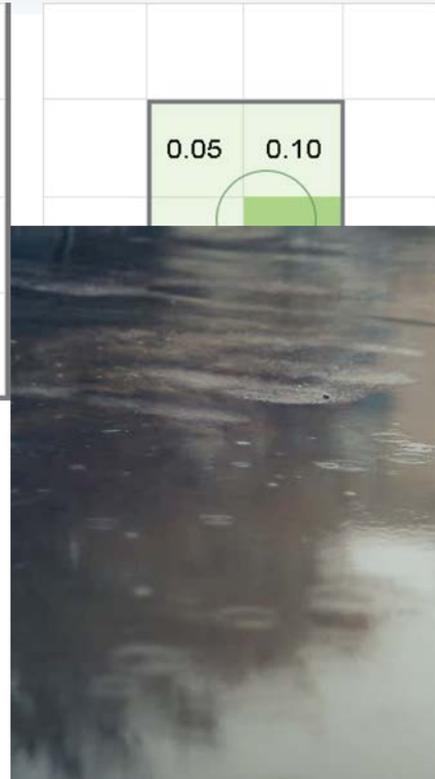
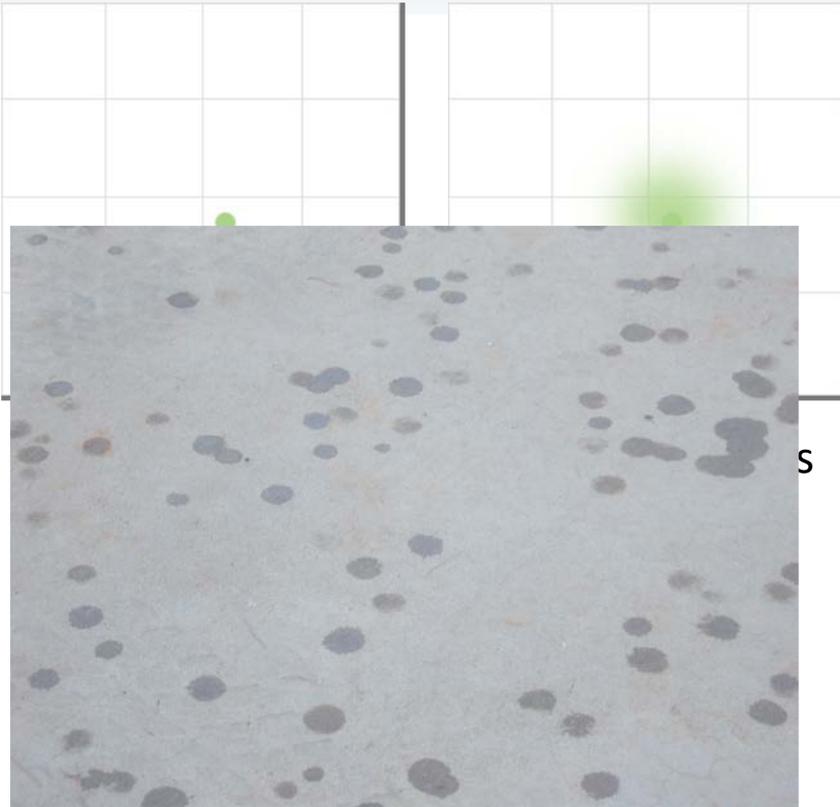


Complementary metal-oxide semiconductor (CMOS)

Integration vs Counting

Integration

- Short exposures
- High Dose-rates



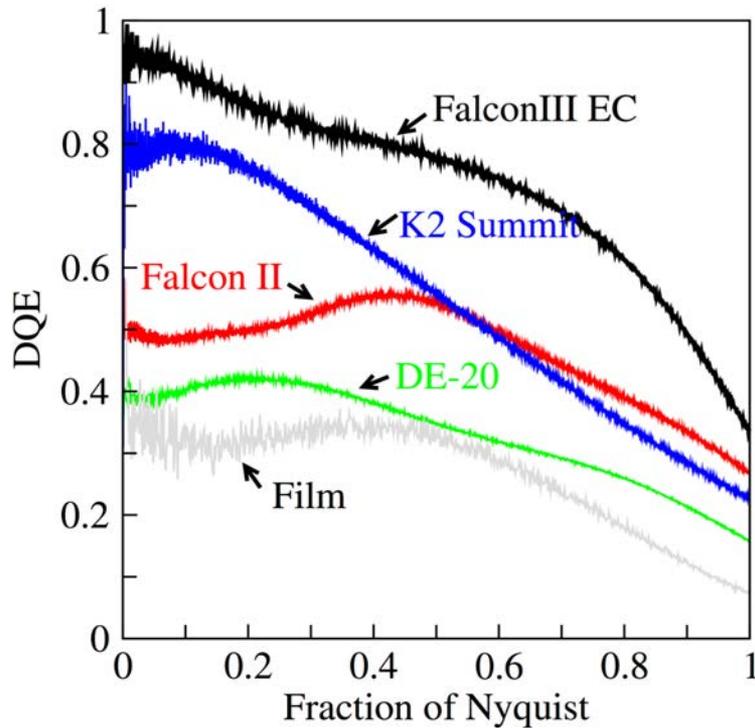
Events reduced to
highest charge pixels.

(0.5-15 e-/pixel/sec)

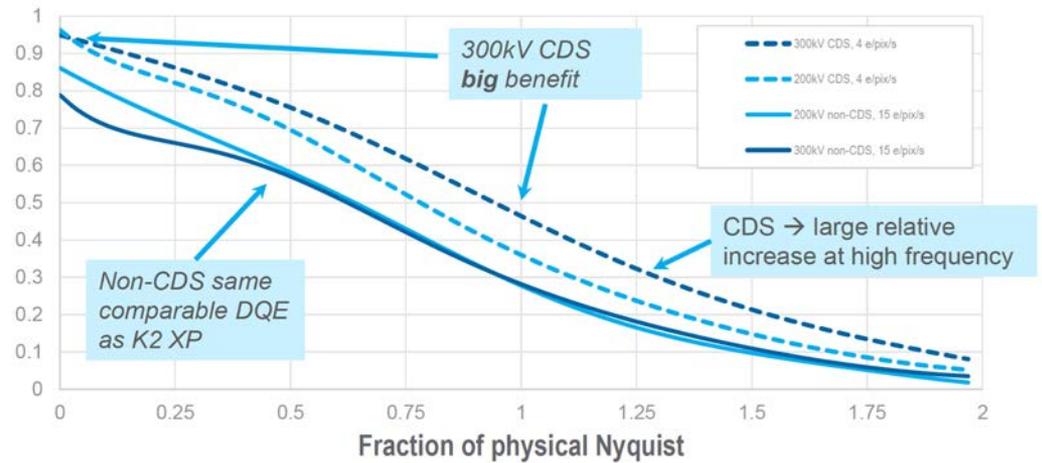
- Fast frame rates
- Long exposures
- Higher DQE

DQE comparison of some detectors

Previous Generation DDD



The GATAN K3 (Paul Mooney, GATAN)

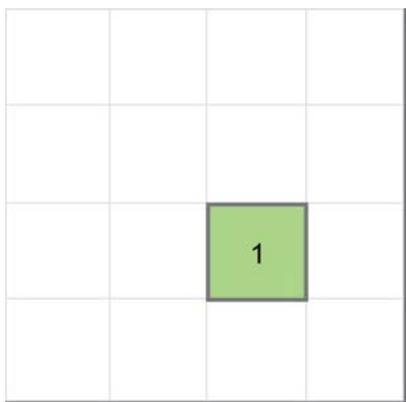


McMullan et. al.
Ultramicroscopy, 2014

Nyquist: 2X the magnified pixel size

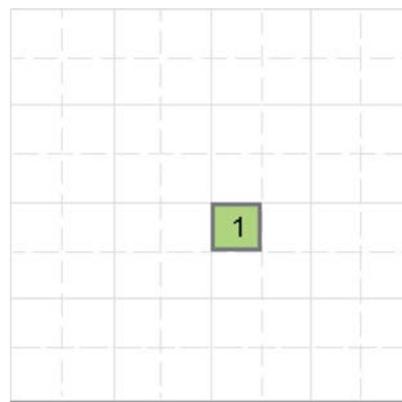
Super-resolution

Counting



Events reduced to
highest charge pixels.

Counting with Super-resolution

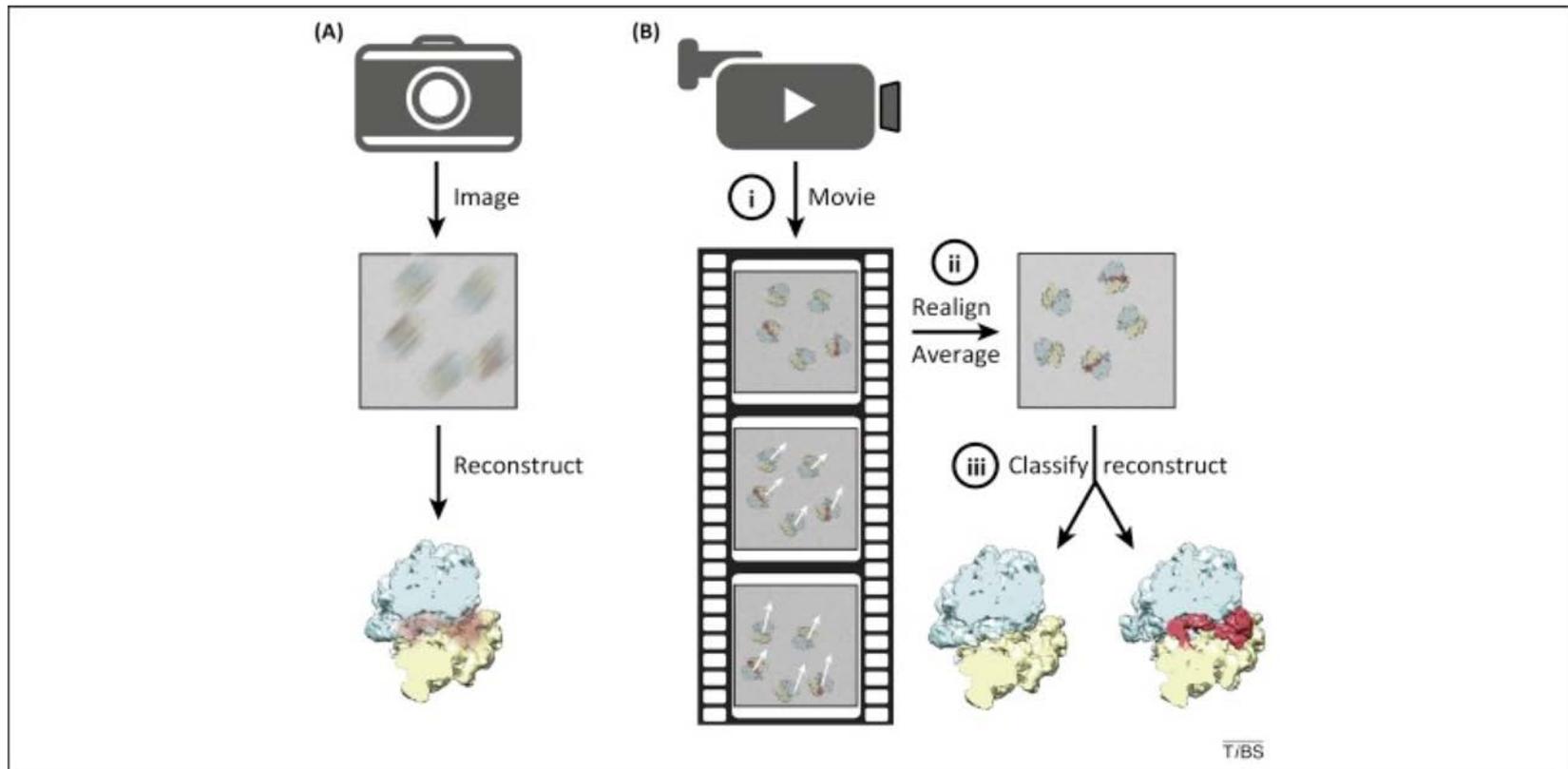


Events Events
localised to sub-pixel
accuracy.

- Super-resolution with Fourier cropping “binning” increases DQE
- Increases disk space requirement
- Even allows one to go beyond physical Nyquist:
 - Recent example at our facility: Data at 81K (Pixel 1.09Å). Collected at SR bin1
 - Resolution reached physical Nyquist (~2.2Å)
 - Re-extracted SR movies during polishing step > Reached 1.9Å

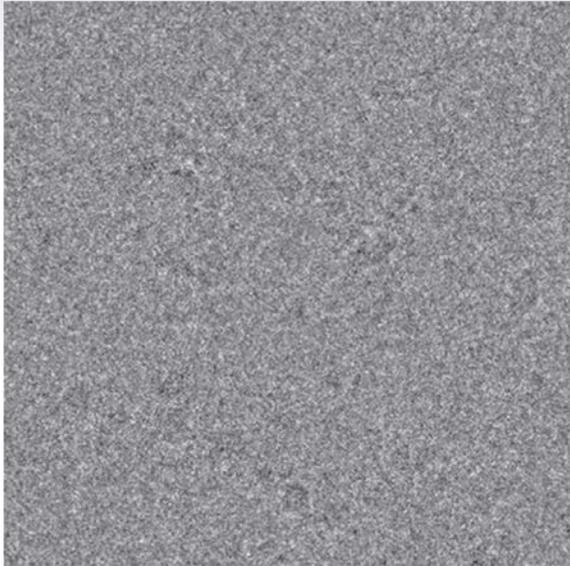
Movies instead of snapshots

- CCD and film limited to one exposure
- Fast frame rate of DED allows movie collection

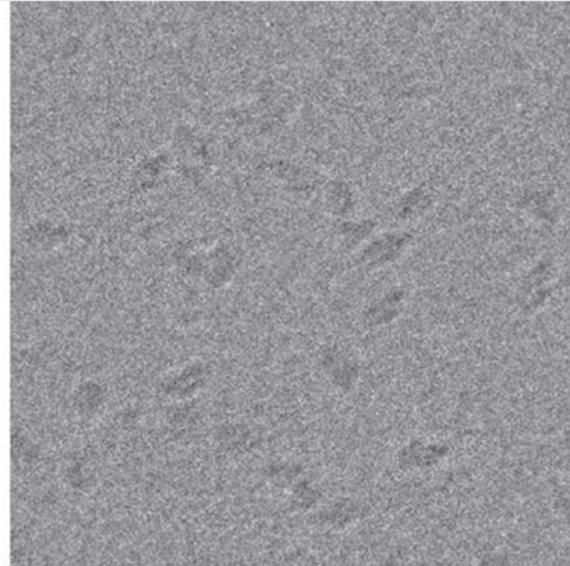


Movies instead of snapshots

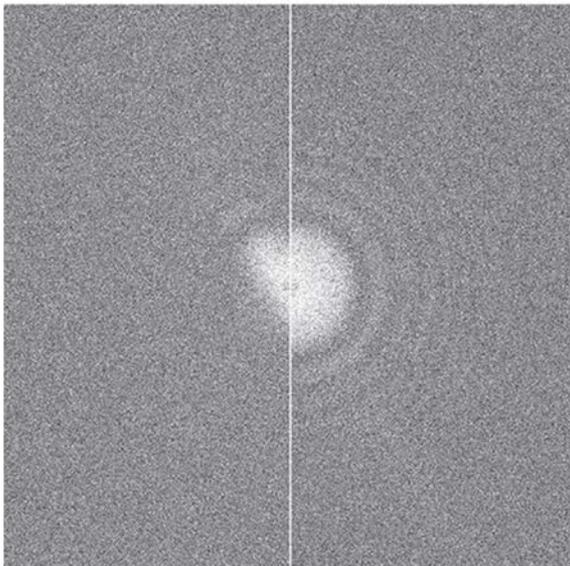
Single Frame



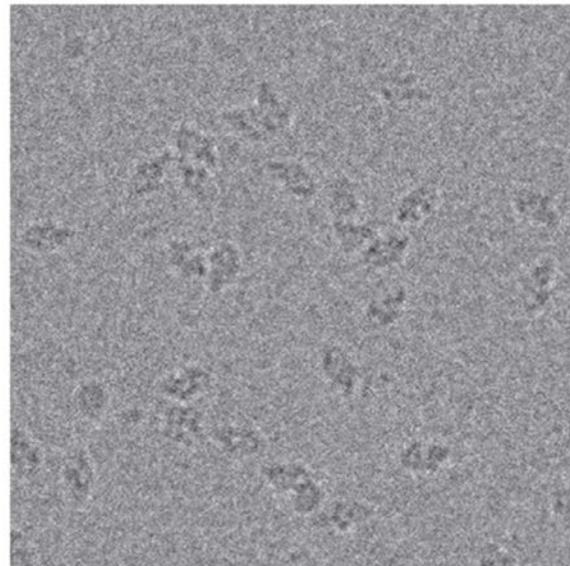
Summed frames
(No alignment)



Power Spectra
Unaligned / Aligned

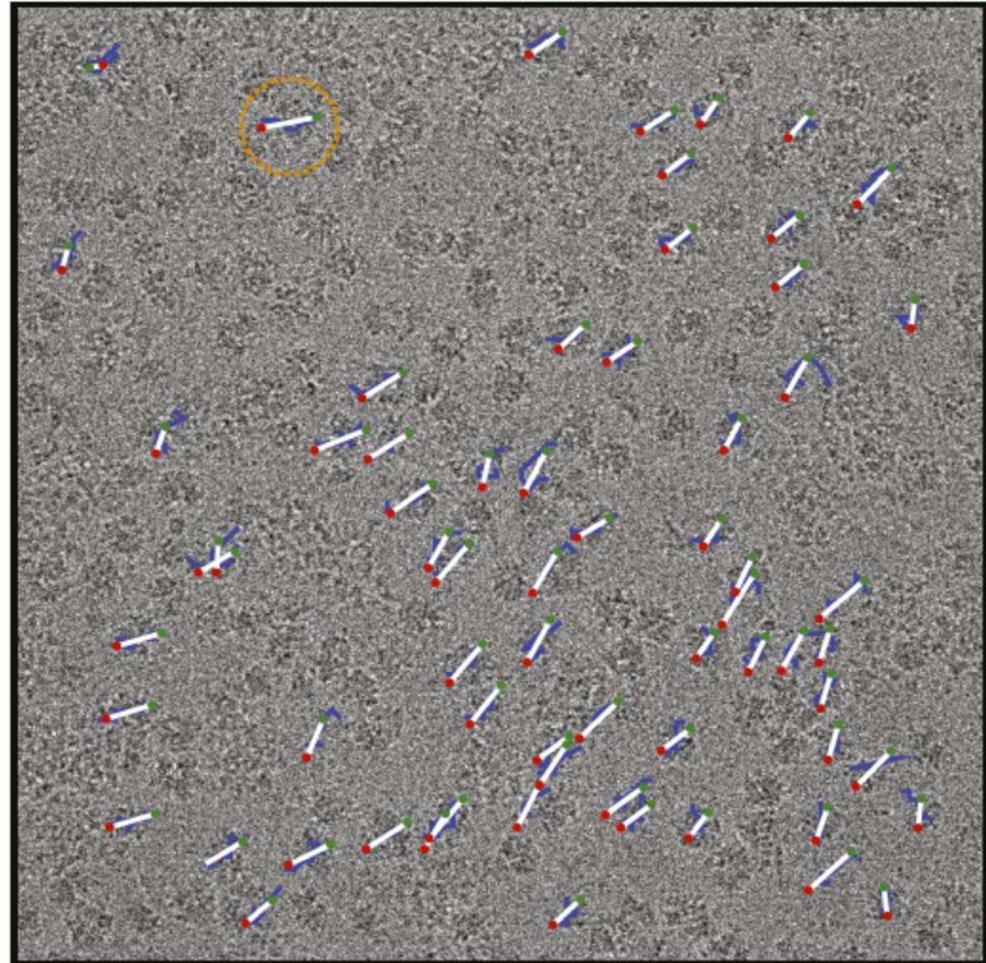


Summed frames
(With alignment)



Individual particle tracking

- Particles can move in the ice
 - Electrostatic attraction
 - Release of stress in the ice

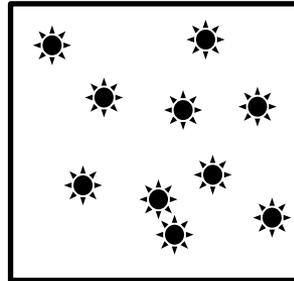


Radiation Damage

- Biological samples are radiation sensitive
- Bonds are broken and free radicals released
- Imaging performed using “Low Dose” methods
- Each micrograph receives a limited amount of electrons to prevent structure deformation
- Typically we use 40-60 electrons/Å² per micrograph

Using movies to deal with radiation damage

“Old days” (before 2012)
 Single image
 1 second exposure at 40
 $e^-/\text{\AA}^2/\text{sec}$ dose rate

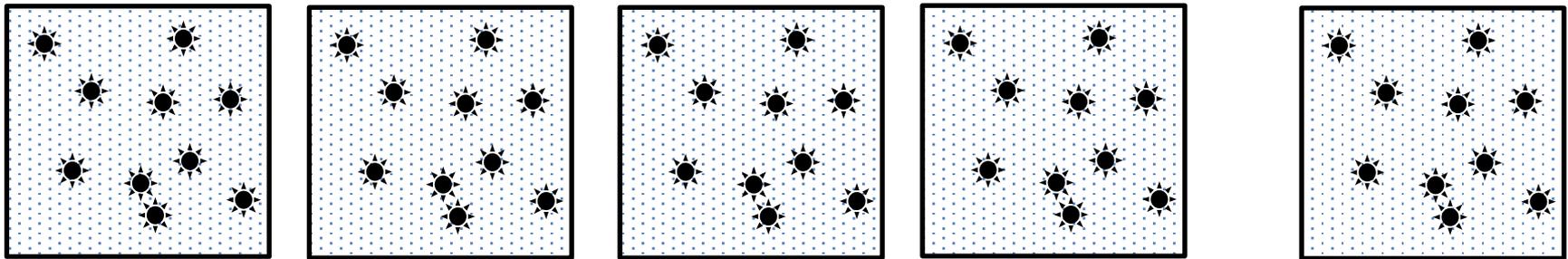


Dose per image: $40 e^-/\text{\AA}^2$

Current date: New detectors
 40 frames/sec, 1 second exposure at $40 e^-/\text{\AA}^2/\text{sec}$ dose rate

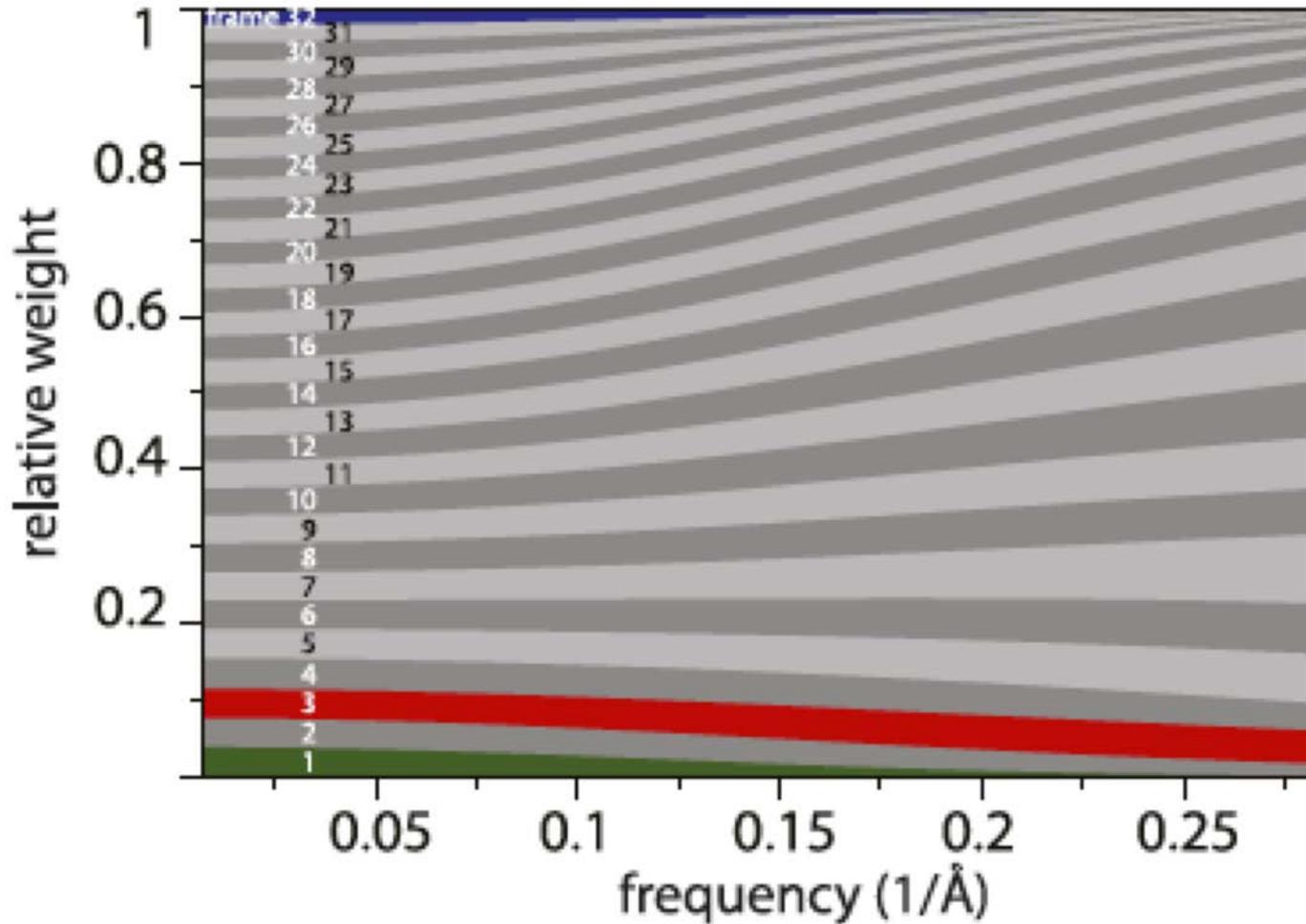
Lower radiation damage: High frequencies are signal

Higher radiation damage: High frequencies are noise



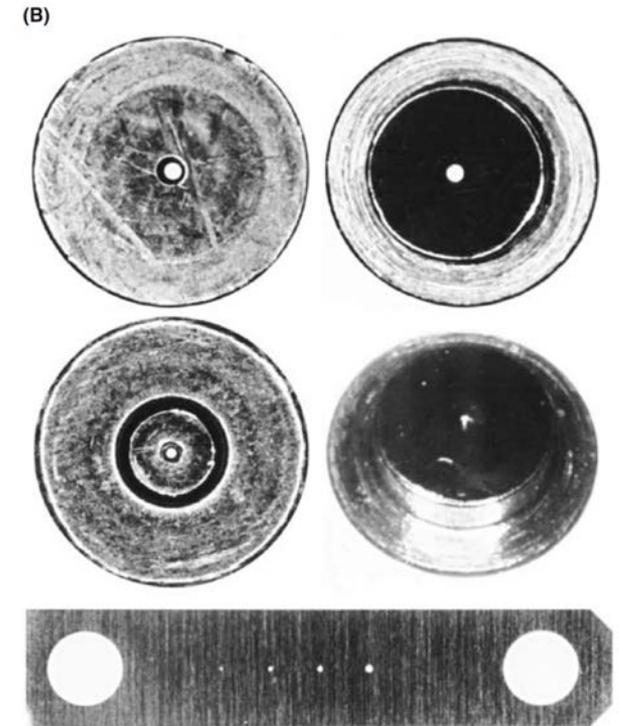
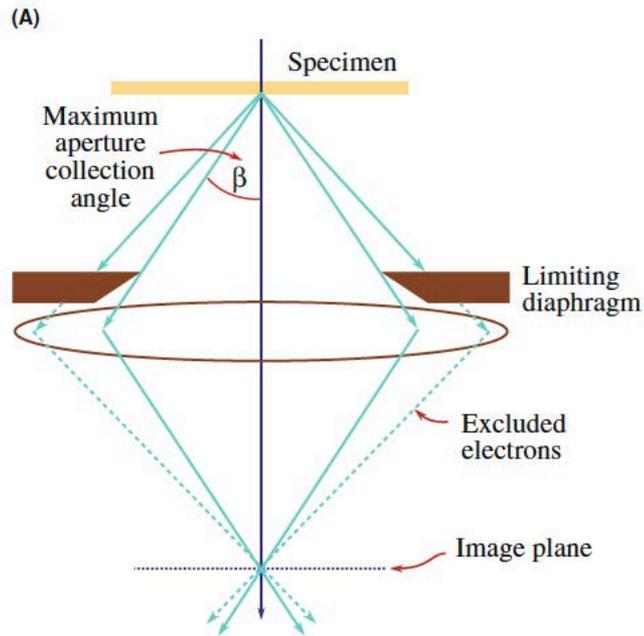
Frame	1	2	3	4	40
Accumulated					
Dose ($e^-/\text{\AA}^2$)	1	2	3	4	40

Radiation damage weighting

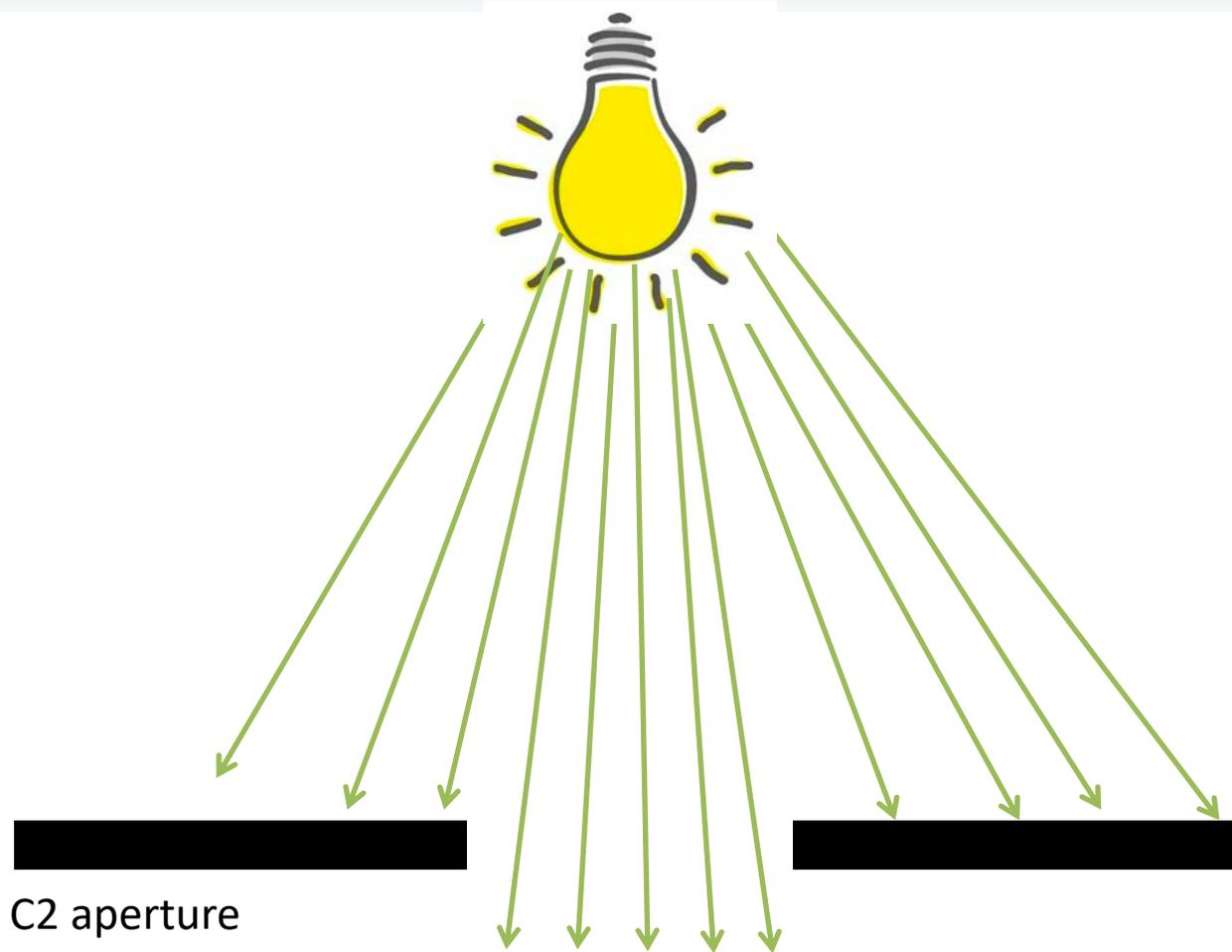


Microscope Apertures

- An aperture is a small hole in a strip of metal inserted in the beam path
- Will only allow the central beam to go through

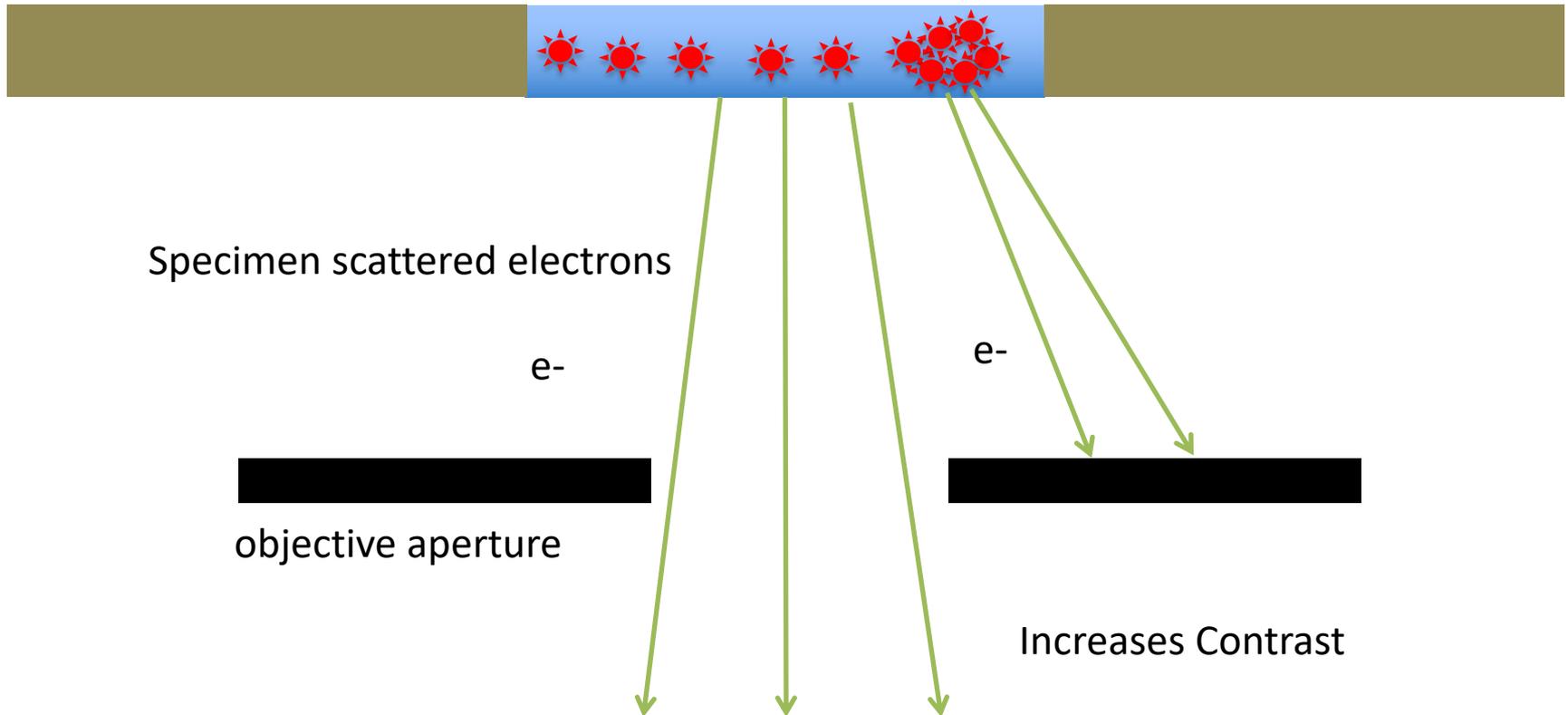


Condenser Aperture



C2 aperture Produces a more coherent, parallel beam

Objective Aperture



Objective Aperture

Also acts as a low-pass filter removing high-resolution information

Consider resolution cut-off. e.g on Krios:

$$30 \mu\text{m} > 4 \text{ \AA}$$

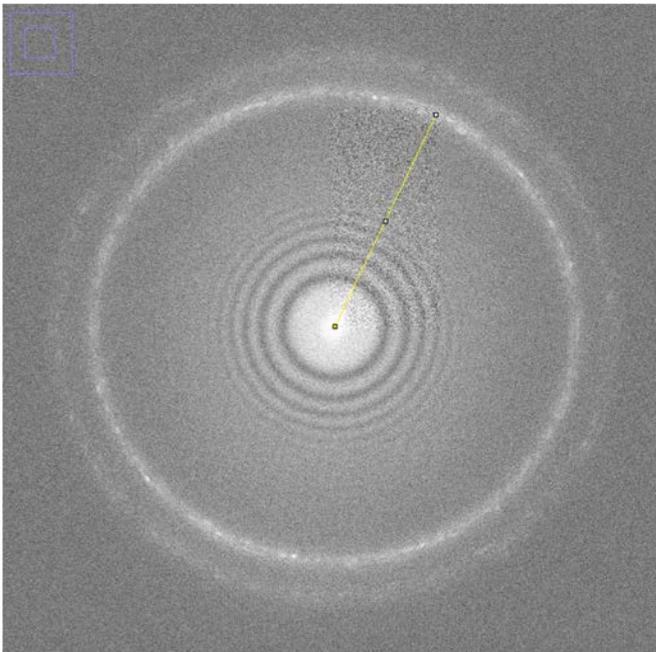
$$70 \mu\text{m} > 1.8 \text{ \AA}$$

$$100 \mu\text{m} > 1.4 \text{ \AA}$$

Magnification and Pixel sizes

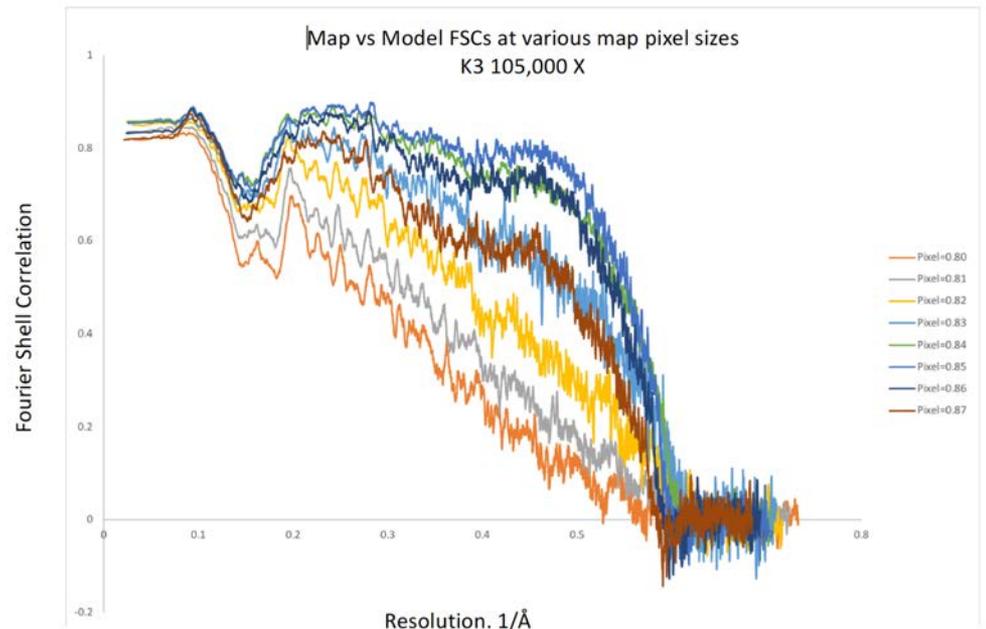
Nominal Magnification

SA 105 kx
nP EFTEM

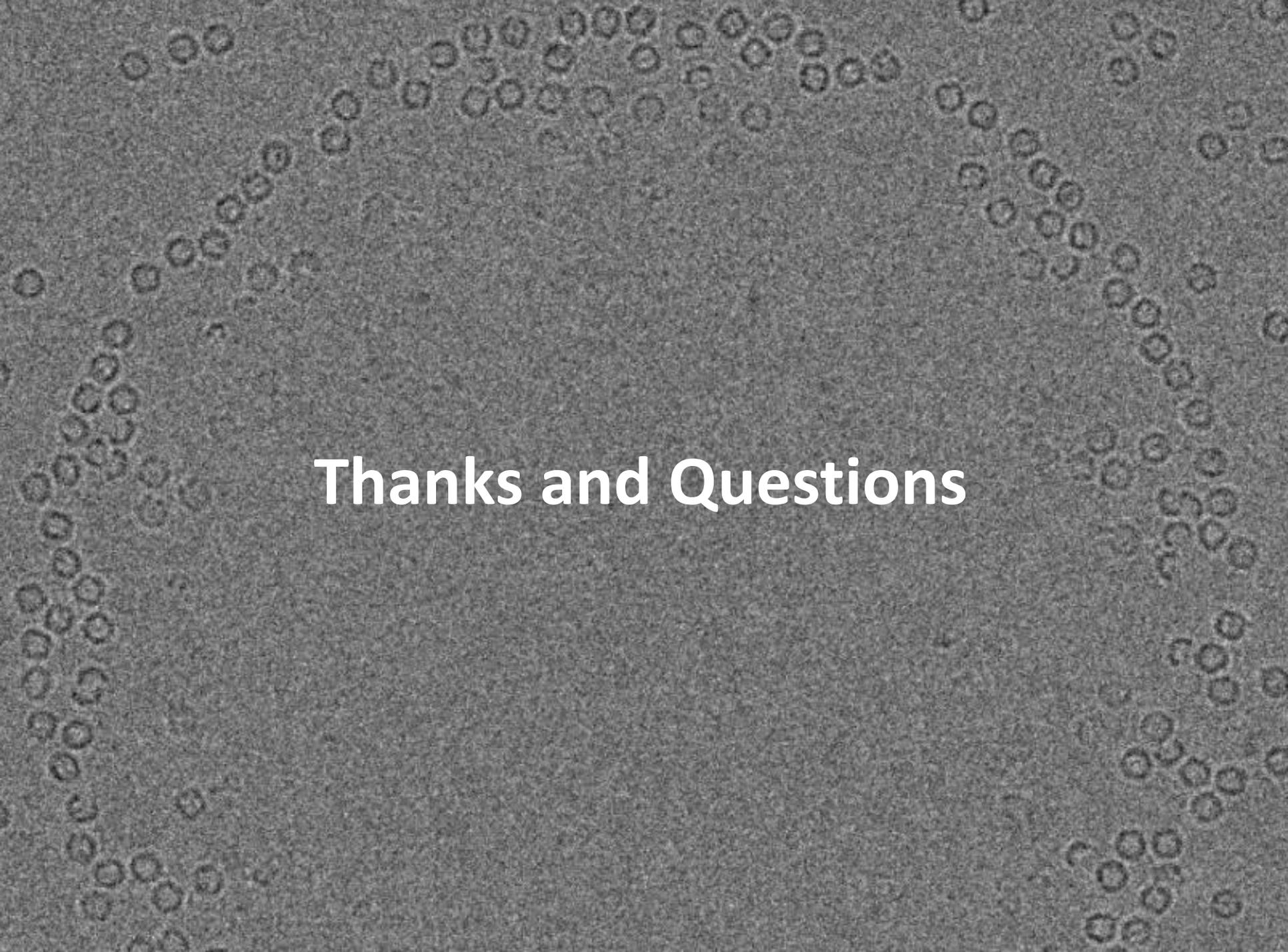


Gold diffraction: 2.35Å

$$\text{Pixel size} = \frac{\text{Calibrated Magnification}}{\text{Physical Pixel Size}}$$



Known atomic model fitting

The background of the slide is a grayscale micrograph showing a dense field of small, circular cells, likely yeast or bacteria, arranged in various patterns and clusters. The cells are slightly out of focus, creating a textured, grainy appearance.

Thanks and Questions